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1 Inflammasomes in the lung

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20

21 **Abstract**

22 Innate immune responses act as first line defences upon exposure to noxious stimuli. The
23 innate immune system has evolved numerous intracellular and extracellular receptors that
24 undertake surveillance for noxious stimuli. Inflammasomes are intracellular innate immune
25 multiprotein complexes that form and are activated following interaction with these stimuli.
26 Inflammasome activation leads to the cleavage of pro-IL-1 β and release of the pro-
27 inflammatory cytokine, IL-1 β , which initiates acute phase pro-inflammatory responses, and
28 other responses are also involved (IL-18, pyroptosis). However, excessive activation of
29 inflammasomes can result in chronic inflammation, which has been implicated in a range of
30 chronic inflammatory diseases. The airways are constantly exposed to a wide variety of
31 stimuli. Inflammasome activation and downstream responses clears these stimuli. However,
32 excessive activation may drive the pathogenesis of chronic respiratory diseases such as severe
33 asthma and chronic obstructive pulmonary disease. Thus, there is currently intense interest
34 in the role of inflammasomes in chronic inflammatory lung diseases and in their potential for
35 therapeutic targeting. Here we review the known associations between inflammasome-
36 mediated responses and the development and exacerbation of chronic lung diseases.

37

38 **Abstract word length: 177**

39 **Keywords:** Inflammasome, asthma, chronic obstructive pulmonary disease, IL-1 β , lung

40 **List of non-standard abbreviations:**

41	AAD	Allergic airways disease
42	AHR	Airways hyperresponsiveness
43	AIM	Absent in melanoma
44	ALI	Acute lung injury
45	Alum	Aluminium hydroxide
46	ARDS	Acute respiratory distress syndrome
47	BALF	Bronchoalveolar lavage fluid
48	CAP	Community acquired pneumonia
49	CAPS	Cryopyrin-associated periodic syndrome
50	Casp1	Caspase-1
51	CARD	Caspase-recruitment domain
52	CF	Cystic fibrosis
53	CFTR	CF transmembrane conductance regulator
54	COPD	Chronic obstructive pulmonary disease
55	DAMP	Damage-associated molecular patterns
56	HDM	House dust mite
57	HIN	Haemopoietic IFN-inducible nuclear
58	HMGB1	Chromatin-binding high motility group box 1 protein
59	IAV	Influenza A virus
60	LRR	C-terminal leucine rich repeats
61	NBD	Nucleotide binding domain
62	NLR	NOD-like receptor
63	NOD	Nucleotide oligomerisation domain
64	NLRC	NOD-like receptor containing
65	NLRP	NOD-like receptor protein

66	Ova	Ovalbumin
67	PAMP	Pathogen-associated molecular pattern
68	PRR	Pattern recognition receptor
69	PYD	Pyrin domain
70	PYHIN	Pyrin and haemopoietic IFN-inducible nuclear
71	ROS	Reactive oxygen species
72	SSI	Severe, steroid-insensitive
73	SLE	Systemic lupus erythematosus
74	TRAF	TNF receptor-associated factor
75	TXNIP	Thioredoxin-interacting protein
76		

77 **1. Introduction**

78 The mucosal surface of the lung is continuously exposed to noxious stimuli that can
79 activate host immunity. As a result, the immune system needs to strike a delicate balance
80 between immune-mediated clearance of these stimuli, and avoiding inadvertent self-harm
81 from chronic inflammation. The term 'noxious stimuli' collectively refers to any stimulus that
82 is capable of causing a tissue-damaging event, such as infection and environmental exposures
83 (e.g. allergens, smoke, pollution) (dos Santos et al., 2012; Medzhitov, 2008). The innate
84 immune system is an evolutionarily conserved, non-antigen specific system that is essential
85 for inducing and orchestrating acute inflammatory responses to infection. In the lung, innate
86 immune responses exert these effects through downstream signalling from numerous
87 germline-encoded pattern recognition receptors (PRRs), which are constitutively expressed
88 by airway epithelial cells, alveolar macrophages, antigen presenting cells and neutrophils (dos
89 Santos et al., 2012; Hallstrand et al., 2014; Medzhitov, 2008). Cell surface as well as cytosolic
90 PRRs, such as Toll-Like Receptors (TLR)s and C-type lectins, continuously survey the
91 extracellular milieu and intracellular compartment, respectively, for pathogen-associated
92 molecular patterns (PAMPs).

93 PAMP-recognising PRRs are split into various families based on their specificity, function
94 and localisation. PRRs can also detect host derived damage-associated molecular patterns
95 (DAMP)s. PRR localisation is important in determining their function, and membrane bound
96 receptors such as Toll-like receptors (TLR)s recognise exogenous extracellular PAMPs or
97 DAMPs. Cytosolic PRRs receptors include the interferon-inducible pyrin and haemopoietic
98 interferon (IFN)-inducible nuclear (PYHIN) family of proteins such as absent in melanoma
99 (AIM)2, which activate type 1 IFN responses, as well as the nucleotide oligomerisation domain
100 (NOD)-like receptors (NLRs) that all recognise PAMPs or DAMPs intracellularly. The
101 recognition of noxious stimuli by these PRRs leads to the induction of innate immune
102 responses.

103 The NLR family recognise a variety of PAMPs as well as host-derived DAMPs (Becker and
104 O'Neill, 2007; dos Santos et al., 2012; Dowling and O'Neill, 2012; Schroder and Tschopp,
105 2010). Recent studies have shown that several NLRs, namely NLR protein (NLRP)1, NLRP3 and
106 NLR family caspase recruitment domain (CARD)-containing (NLRC4) can assemble into multi-
107 protein complexes termed inflammasomes that recruit and activate pro-inflammatory
108 caspases, such as Caspase-1 (Casp1). Inflammasome-activated Casp1 in turn cleaves pre-

109 cursor forms of pro-inflammatory cytokines in the interleukin (IL)-1 family to produce active
110 IL-1 β and IL-18, and inactivates IL-33 (Martinon et al., 2002; Martinon et al., 2009; Schroder
111 and Tschopp, 2010; Stutz et al., 2009).

112 To date, five inflammasomes have been studied in detail; the NLR family members, NLRP1,
113 NLRP3, NLRC4, and retinoic acid-inducible gene 1 (RIG-I), and AIM2. Following assembly and
114 activation, all of these inflammasomes can enzymatically cleave and activate Casp1 in
115 response to DAMP signalling (Dowling and O'Neill, 2012).

116 Clinical and experimental evidence increasingly and strongly implicates excessive
117 inflammasome activation and production of IL-1 β in the pathogenesis of several chronic
118 respiratory diseases, including severe, steroid-insensitive (SSI) asthma and chronic
119 obstructive pulmonary disease (COPD) (Aaron et al., 2001; Baines et al., 2011; Chung, 2001;
120 Hastie et al., 2010; Kim et al., 2014a; Kim et al., 2015; Konno et al., 1996; Simpson et al.,
121 2014a; Wanderer, 2000). Here we review the current literature on the association between
122 inflammasome-mediated host innate immune responses and the development and
123 exacerbation of chronic lung diseases.

124

125 **1.1 IL-1 β activation in the lung**

126 Through respiration the airway lumen is continually exposed to potentially pathogenic
127 micro-organisms (viral, fungal and bacterial) and environmental stimuli (exogenous noxious
128 pollutants such as cigarette, cooking or wood smoke, air pollution, particulate matter, silica,
129 asbestos or elemental metals). The homeostatic regulation of immune responses, and the
130 ability to distinguish self from noxious stimuli, involves complex communication and inter-
131 regulation between the innate and adaptive immune systems. Innate immunity relies on the
132 activation of germline-encoded PRRs to mount immediate responses to infectious and
133 noxious stimuli. Epithelial cells and activated macrophages and neutrophils are the first cells
134 to respond following PRR activation. They play critical roles in mediating the adaptive immune
135 response through the release of cytokines that promote the chemo-attraction of adaptive
136 immune cells. Thus, innate immune responses have significant bearing on the ensuing
137 adaptive immune response and, if unregulated, can result in severe pathological
138 consequences (Hirota and Knight, 2012; Schroder and Tschopp, 2010; Stutz et al., 2009; Yang
139 et al., 2012).

140 The NLRs are an important family of intracellular PRRs that can be activated by PAMPs
141 such as lipopolysaccharide (LPS) or DAMPs including those of endogenous origin, such as
142 adenosine tri-phosphate (ATP) and uric acid crystals that are released from lysed cells, and/or
143 DAMPs of exogenous origin, including inhaled silica, the adjuvant aluminium hydroxide (alum)
144 and smoke, particulate matter and allergens (Eisenbarth et al., 2008; Hirota et al., 2012;
145 Hornung et al., 2008; Shi et al., 2003). The NLR family is comprised of 22 members that are
146 characterised based on their C-terminal leucine rich repeats (LRR) domains and central NACHT
147 nucleotide-binding domains (NBD). The LRR domains detect NLR-specific ligands and, in the
148 absence of ligand-mediated activation, play an important role in the auto-inhibitory
149 regulation of inflammasomes. NLR family members are also categorised based on their N-
150 terminal domains and the largest group, containing 14 members, possesses distinct N-
151 terminal pyrin domains (PYD) (Dowling and O'Neill, 2012; Martinon et al., 2002; Schroder and
152 Tschopp, 2010; Stutz et al., 2009). To date 14 unique NLRP family members (termed NLRP1-
153 14) have been identified and classified (Zhang et al., 2008). Some NLRs possess a common N-
154 terminal CARD (e.g. NLRC4) and variations of NODs (e.g. NOD1 [NLRC1] and NOD2 [NLRC2])
155 (Dowling and O'Neill, 2012; Martinon et al., 2002; Schroder and Tschopp, 2010; Stutz et al.,
156 2009). All the members of the NLR family have critical roles in the detection of noxious stimuli
157 in the cytosol and, following ligation, induce the proteolytic cleavage and activation of IL-1 β .

158 IL-1 β is a potent, pyrogenic, pro-inflammatory cytokine that plays important roles in
159 initiating acute inflammatory responses to infectious, noxious or cell damage-derived stimuli.
160 In the lung, activation of PRRs such as TLR2 or TLR4 induces the transcription of biologically
161 inactive pro-IL-1 β . Pro-IL-1 β is primarily produced by macrophages but is also generated in
162 neutrophils, lymphocytes, airway epithelial cells and fibroblasts (Dinarello, 1994). Proteolytic
163 cleavage of pro-IL-1 β generates bio-active IL-1 β that is then released into the extracellular
164 milieu and exerts its pro-inflammatory effects by binding to the extracellular domain of the
165 ubiquitously expressed IL-1 receptor, type 1 (IL-1R)-1. Ligand-receptor interactions result in
166 the recruitment of the secondary receptor chain IL-1Racp (Sims et al., 1988). This initiates the
167 formation of a receptor complex that recruits the pro-inflammatory adaptor molecules
168 myeloid differentiation primary response gene (MyD)88, IL-1R-associated kinase 1 (IRAK) and
169 tumour necrosis factor (TNF) receptor-associated factor (TRAF)6 (Greenfeder et al., 1995). In
170 turn, this results in the activation of the pro-inflammatory transcription factor nuclear factor
171 kappa-light-chain-enhancer of activated B cells (Nf- κ B). This induces the release of TNF- α and

172 IL-6, promotes the recruitment and activation of innate immune cells such as neutrophils and
173 macrophages, and further expression of pro-IL-1 β (Greenfeder et al., 1995). Since active IL-1 β
174 is secreted into the extracellular milieu it can exert pro-inflammatory effects through both
175 autocrine and paracrine mechanisms and, depending on the circulating levels, can lead to
176 severe systemic inflammation (Attur et al., 2000).

177 Activation and release of IL-1 β plays a major role in acute inflammatory responses to
178 respiratory infection and promotes clearance of pathogens. However, prolonged and
179 unmitigated activation and release can lead to chronic systemic inflammation such as that
180 observed in Muckle-Wells syndrome (also known as cryopyrin-associated periodic syndrome
181 [CAPS]) where a gain-of-function mutation in the NLRP3 gene leads to chronic activation of
182 IL-1 β causing fever, skin rash, joint pain, and conjunctivitis (Broderick et al., 2015; Coll et al.,
183 2015). Under normal conditions, the activation and release of IL-1 β occurs in a tightly
184 controlled two-stage process that requires an initial priming step provided by PAMP-
185 mediated activation of PRRs (e.g. TLR4 ligation following exposure to LPS), and a secondary
186 activating signal provided by DAMPs (e.g. ATP) (dos Santos et al., 2012; Schroder and Tschopp,
187 2010; Takeda et al., 2003). This causes inflammasome assembly and activation and leads to
188 the proteolytic cleavage and activation of the protease zymogen, pro-Casp1, which was
189 originally described as interleukin-converting enzyme (ICE) (Thornberry et al., 1992).
190 Intriguingly, recent research investigating the therapeutic potential of targeting this
191 inflammasome–Casp1–IL-1 β axis has focused on the development of monoclonal antibodies
192 that target IL-1 β and/or IL-1R. Anakinra, an IL-1R antagonist, has been trialled in patients with
193 chronic rheumatoid arthritis, however, a study by Listing *et al.*, showed that treatment
194 increased the risk of infection compared to conventional therapy with disease modifying anti-
195 rheumatic drugs (Listing et al., 2005).

196 Excess activation of the inflammasome can also lead to pyroptosis, which is a form of
197 rapid lytic cell death. Sagulenko *et al.*, identified that dsDNA, another DAMP, increases the
198 activation of the AIM2 inflammasome in a dose-dependent manner, leading to pyroptosis
199 (Sagulenko et al., 2013). It is therefore important that inflammasome responses are tightly
200 regulated in order to prevent the development of chronic inflammatory diseases.

201

202 **2. The NLRP3 inflammasome in the lung**

203 NLRP3 is the best characterised and most widely implicated inflammasome in
204 inflammatory diseases of the lung. Activation of the NLRP3 inflammasome complex is
205 important for combating respiratory infections, however, excessive activation may contribute
206 to the development of severe disease. It is comprised of an NLRP3 domain that possesses a
207 PYD central NBD and C-terminal LRRs (O'Connor et al., 2003), and an adaptor protein,
208 apoptosis-associated speck-like protein containing a CARD (ASC), which recruits and activates
209 pro-Casp1 (Mariathasan et al., 2004; Masumoto et al., 1999). Active Casp1 subsequently
210 cleaves pro-IL-1 β and pro-IL-18 to their active secreted forms (Mariathasan et al., 2004).

211 In the lung, the NLRP3 inflammasome can be activated by a diverse range of microbial
212 pathogens, including opportunistic bacteria (*Klebsiella pneumoniae*, *Streptococcus*
213 *pneumoniae* and other spp. and *Haemophilus influenzae* (Mariathasan and Monack, 2007;
214 Rotta Detto Loria et al., 2013; Willingham et al., 2009)), atypical bacteria (*Chlamydia* (Abdul-
215 Sater et al., 2010)) and viruses (including influenza A virus (IAV), vesicular stomatitis virus
216 (VSV) and adenovirus (AdV)) (Table 1) (Muruve et al., 2008; Rajan et al., 2011; Wang et al.,
217 2014). A study by Costa *et al.*, showed that NLRP3-deficient mice are more susceptible to
218 group B *Streptococci* respiratory infection than wild-type mice (Costa et al., 2012).
219 Furthermore, *Streptococcus*-derived pneumolysin has been shown to induce NLRP3
220 inflammasome-mediated responses and downstream IL-17A and IFN- γ responses to confer
221 protection against infection (McNeela et al., 2010). These studies suggest that the NLRP3
222 inflammasome plays a critical role in the clearance of respiratory infections. Furthermore, a
223 recent study by Tate *et al.*, examined the temporal effect of NLRP3 inflammasome inhibition
224 using the novel, highly specific NLRP3 inflammasome inhibitor, MCC950, in IAV infected mice.
225 They showed that inhibition of NLRP3 in the early phase of infection led to hyper-susceptibility
226 to lethality, whereas inhibition of NLRP3 at the peak of infection significantly protected mice
227 from severe disease (Tate et al., 2016). These data highlight that NLRP3 activation may be
228 beneficial in acute phase inflammation in IAV infection, but may be detrimental in more
229 chronic inflammatory disease.

230 Inhaled environmental stimuli (such as smoke, asbestos, silica, metal alloys and
231 particulate matter and air pollution), are also capable of activating the NLRP3 inflammasome
232 (Hirota et al., 2015; Hirota et al., 2012; Kim et al., 2015), and several studies have linked
233 exposure to airway pollutants with the development of chronic airway diseases. A recent
234 study by Hirota *et al.*, showed that particulate matter derived from airway pollutants

235 collected from developed areas can activate the NLRP3 inflammasome *in vitro* (Hirota et al.,
236 2012). This study showed that particulate matter up to 10 μ M in diameter (PM₁₀) activated
237 the NLRP3 inflammasome in epithelial cells. The same team also reported that this could
238 occur through the induction of innate immune responses (Hirota et al., 2015). Other studies
239 have shown that endogenous DAMPs, such as extracellular ATP, can also activate the NLRP3
240 inflammasome (Mariathasan et al., 2006; Zhou et al., 2011). The presence of DAMPs in the
241 airway is a strong indication of prior pathogenic insult resulting in cell lysis and/or chronic
242 inflammation. Aeffner *et al.*, and others, recently showed that IAV, which infects the
243 respiratory epithelium and is a known inflammasome activator, causes the release of ATP into
244 the airway lumen, potentially highlighting the mechanism by which this virus activates the
245 NLRP3 inflammasome (Aeffner et al., 2011; Allen et al., 2009; Muruve et al., 2008). Several
246 studies have shown that ATP and K⁺ efflux activates the NLRP3 inflammasome through the
247 P2X₇ purinoceptor 7 (P2X₇R), which is a cation-permeable ligand-gated ion channel. ATP-
248 mediated activation results in P2X₇R pore formation of the pannexin-1 hemi-channel that
249 then facilitates passive migration of extracellular PAMPs and DAMPs into the cytosol where
250 they can induce the assembly and activation of the NLRP3 inflammasome (Chaudhuri et al.,
251 2010; Kanneganti et al., 2007).

252 Recent studies have also highlighted a potential role for oxidative stress, induced by the
253 production of reactive oxygen species (ROS), in promoting the assembly and activation of the
254 NLRP3 inflammasome. Zhou *et al.*, showed that mitochondria-derived ROS (mtROS) induces
255 the activation of the NLRP3 inflammasome in the endoplasmic reticulum (Zhou et al., 2011).
256 They also showed that mtROS contributes to the assembly of the NLRP3 inflammasome *via*
257 the thioredoxin and thioredoxin-interacting (TXNIP) complex. ROS interact with the TXNIP
258 complex and cause it to dissociate from thioredoxin. This allows dissociated TXNIP to bind
259 with NLRP3 that then recruits ASC and Casp1 to form the inflammasome complex (Zhou et al.,
260 2010). It remains possible that one mechanism by which air pollution activates the
261 inflammasome in the lung is via the oxidative capacity of the components of particulate
262 matter (e.g. elemental metals and poly aromatic hydrocarbons). Collectively, these data
263 suggest that exogenously-induced oxidative stress and ROS and can also induce the assembly
264 of the NLRP3 inflammasome.

265

266 **3. The AIM2 inflammasome and the lung**

267 The AIM2 inflammasome belongs to the PYHIN family of proteins and is comprised of an
268 N-terminal PYD domain and one or two copies of a 200 amino acid repeat haemopoietic IFN-
269 inducible nuclear proteins (HIN) domain at its C-terminus (Dowling and O'Neill, 2012). The HIN
270 motifs detect DNA of microbial origin and induce type 1 IFN responses and the expression of
271 pro-IL-1 β (Dowling and O'Neill, 2012).

272 Several studies have demonstrated that exposure to microbial or mammalian dsDNA
273 leads to Casp1 activation in an AIM2-dependent manner (Fernandes-Alnemri et al., 2010;
274 Hornung et al., 2009; Muruve et al., 2008; Roberts et al., 2009; Schroder et al., 2009) (Table
275 1). Cytosolic dsDNA is detected by the HIN-200 domain of the AIM2 inflammasome that then,
276 through its PYHIN domain, recruits ASC and pro-Casp1. This results in the proteolytic cleavage
277 and activation of Casp1 and IL-1 β (Hornung et al., 2009; Muruve et al., 2008; Roberts et al.,
278 2009; Schroder et al., 2009). Fernades-Almenri *et al.*, recently showed that AIM2-deficient
279 mice exhibit increased mortality following *Francisella tularensis* respiratory infection
280 compared to wild-type controls, which indicated that AIM2 inflammasome-mediated
281 responses are required for effective clearance of infection. Since *F. tularensis* replicates in the
282 cytosol of infected macrophages the authors concluded that pathogen-derived DNA is
283 detected by the AIM2 inflammasome during the process of its escape (Fernandes-Alnemri et
284 al., 2010). Collectively, these data suggest that AIM2 inflammasome-mediated responses are
285 important in immune responses to microbial pathogens that produce dsDNA as part of their
286 replication strategy, and highlight the need for its further exploration. Given that increased
287 IL-1 β is a feature of many chronic respiratory conditions that are also associated with
288 respiratory infections, the further examination of the AIM2 inflammasome in these disease
289 contexts is warranted to elucidate whether it plays a pathogenic role in driving disease.

290

291 **4. Other inflammasomes in the lung**

292 Our understanding of the roles of inflammasomes in the development of chronic lung
293 diseases is in its infancy and research efforts to date have focussed primarily on characterising
294 the roles of the NLRP3 and AIM2 inflammasomes. As a result, the functional roles of other
295 inflammasomes in immune responses to respiratory infections and in the development
296 and/or exacerbation of chronic lung diseases are yet to be resolved. The NLRP1
297 inflammasome was the first to be described and murine studies showed that it is directly
298 activated by anthrax lethal toxin, which is produced by *Bacillus anthracis*, and is able to cleave

309 Casp1 in the absence of ASC (Boyden and Dietrich, 2006; Martinon et al., 2002) (Table 1).
310 Activation of NLRP1 by anthrax lethal toxin is required in mice for the maturation and release
311 of IL-1 β . This results in rapid cell death and is the causative agent of systemic anthrax toxicity
312 (Banks et al., 2006). The development of therapeutics specifically targeting this
313 inflammasome may be crucial in preventing the lethal toxicity of anthrax.

314 The NLRC4 inflammasome, also known as CARD12, has been mostly associated with the
315 induction of pyroptosis (Dowling and O'Neill, 2012). However, in the lung it has been shown
316 to be activated by cytosolic flagellin and the basal rod component of the type 3 secretion
317 system found in a range of bacteria such as *Salmonella typhimurium*, *Shigella flexneri* and
318 *Legionella pneumophila* (Sutterwala et al., 2007).

309

310 **5. Regulation and activation of inflammasomes**

311 The precise mechanisms that lead to NLRP3 inflammasome activation remain poorly
312 defined. However, as mentioned previously this process consists of two distinct events, the
313 first of which involves the expression and assembly of inflammasome components (Schroder
314 and Tschopp, 2010; Stutz et al., 2009). Increasing evidence shows that PAMP-induced TLR
315 ligation and increased levels of TNF- α are important in the initiation of the expression and
316 assembly of NLRP3 inflammasome components as well as inducing pro-IL-1 β expression
317 through the activation of NF- κ B (He et al., 2013; Hiscott et al., 1993; Martinon et al., 2002;
318 Martinon et al., 2009; Schindler et al., 1990a; Schindler et al., 1990b)) (Table 1). The
319 mechanism of NLRP3 activation involves its translocation to mitochondria and the induction
320 of ROS in these organelles, K⁺-efflux and cathepsin release. However, this is an intense area of
321 investigation and recent studies have found new roles for NEK7 and NIMA-related kinase
322 and NIMA-related kinase (He et al., 2016). Interestingly, a study by Ikejima *et al.*,
323 demonstrated that rabbits injected with recombinant IL-1 β produced endogenous IL-1 β ,
324 indicating that IL-1 β responses can self-perpetuate (Ikejima et al., 1990). The authors also
325 showed that human peripheral blood mononuclear cells produced TNF- α in response to
326 treatment with IL-1 β . Thus, in NLRP3 inflammasome-mediated responses, the induction of
327 pro-IL-1 β expression constitutes a form of 'priming' that then requires a secondary round of
328 activation of assembled inflammasome components to drive the proteolytic pathway that
329 leads to the activation and release of IL-1 β (Hornung and Latz, 2010). This represents an
330 important mechanism of immune regulation given that unmitigated activation and release of

331 IL-1 β and other IL-1 family members can have destructive consequences (Dinarello, 2009).
332 Many studies mimic this priming effect by pre-treating cultured cells *in vitro* with TLR agonists
333 and/or pro-inflammatory cytokines, such as LPS and TNF- α (Martinon et al., 2002; Schroder
334 and Tschopp, 2010; Stutz et al., 2009). Recent research has focused on characterising the role
335 of the NLRP3 inflammasome in disease development.

336 There is, however, a paucity of information relating to the role of other inflammasomes
337 in disease and it is important to interrogate these further. *Guarda et al.*, showed that type 1
338 IFNs are capable of suppressing NLRP1 and NLRP3 inflammasome activity, which resulted in
339 diminished Casp1-dependent IL-1 β maturation, suggesting that both of these inflammasomes
340 are equally important in activating IL-1 β (Guarda et al., 2011). Several studies have also
341 examined the role of autophagy, a normal cellular process that deals with the controlled
342 degradation of defective organelles in the cell, and inflammasome activation. *Saitoh et al.*,
343 showed that cells deficient in the autophagy-specific protein Atg16L1 had elevated
344 endotoxin-induced IL-1 β production (Saitoh et al., 2008). Further characterisation of the
345 regulation of the other inflammasomes, such as the AIM2 inflammasome, is essential to
346 increase our understanding of their roles in the pathogenesis of chronic lung diseases. Our
347 understanding of the activation and regulation of inflammasomes is rapidly evolving and
348 therapeutic intervention through numerous avenues could be possible once their biology
349 has been elucidated.

350

351 **6. Inflammasomes in lung diseases**

352 Whilst inflammasomes play crucial roles in the clearance of pathogens, aberrant
353 inflammasome-mediated IL-1 β responses are strongly implicated in the pathogenesis of a
354 wide range of human diseases such as metabolic dysregulation, systemic autoimmune
355 diseases such as systemic lupus erythematosus (SLE), inflammatory diseases of the skin and
356 joints such as rheumatoid arthritis and gout, and chronic inflammatory lung diseases such as
357 asthma and COPD (Baechler et al., 2003; Martinon et al., 2006; Simpson et al., 2014a; Strowig
358 et al., 2012; Yang et al., 2015). Nevertheless, the roles of inflammasomes in the development
359 and exacerbation of lung disease are not well understood and further characterisation of the
360 mechanisms of activation of individual inflammasomes will provide greater insight into their
361 roles. Here we review the existing literature that links aberrant inflammasome activation with
362 the development, exacerbation and progression of chronic airway disease.

363

364 **6.1 Inflammasomes in community acquired pneumonia**

365 Community acquired pneumonia (CAP) refers to any form of pneumonia that is
366 contracted outside a hospital environment. It is a serious life-threatening condition and is the
367 leading cause of infectious disease mortality in many countries (Mizgerd and Skerrett, 2008).
368 Pneumonia is primarily caused by respiratory bacterial infection and results from fluid filling
369 the alveolar spaces. Infection with *S. pneumoniae* is by far the most common cause of CAP (File,
370 2004). A recent study by Witzenrath *et al.*, examined a *S. pneumoniae* infection murine model
371 and determined that the streptococcal exotoxin pneumolysin was essential for NLRP3
372 inflammasome activation (Witzenrath *et al.*, 2011). Pneumolysin disrupts the plasma
373 membrane in cells leading to K⁺ efflux, which could lead to the activation of the NLRP3
374 inflammasome (Schroder and Tschopp, 2010). As described previously, mice deficient in
375 NLRP3 had increased mortality following *S. pneumoniae* infection, therefore the activation of
376 the NLRP3 inflammasome by this bacterium is likely to be essential for the clearance of
377 infection and may be an important factor in CAP (McNeela *et al.*, 2010; Witzenrath *et al.*,
378 2011).

379

380 **6.2 Inflammasomes in asthma**

381 Asthma is a chronic inflammatory disease of the airways that affects approximately 10%
382 of the population in Westernised countries and 300 million sufferers worldwide (Akinbami *et al.*
383 *et al.*, 2012; Bateman *et al.*, 2008; Hansbro *et al.*, 2008; Hansbro *et al.*, 2011; Wang *et al.*, 2010).
384 The majority of asthmatic patients exhibit a mild to moderate form of the disease that is
385 characterised by T-helper lymphocyte type 2-mediated, eosinophil-dominated immune
386 responses (Hansbro *et al.*, 2013; Shahidi and FitzGerald, 2010). Most commonly, asthma is
387 characterised as an allergic disorder and >50% of asthmatics have some form of atopy (Arbes
388 *et al.*, 2007). Increasing evidence now shows that asthma is a heterogeneous disease that is
389 associated with a range of phenotypes and endotypes (Hansbro *et al.*, 2011; Hansbro *et al.*,
390 2013). The majority of asthmatics can control their symptoms through the use of inhaled
391 short-acting β -agonists and inhaled corticosteroids. However, some asthmatics have
392 persistent airflow obstruction, more frequent exacerbations and remain symptomatic despite
393 high doses of these drugs (Bell and Busse, 2013). These asthmatics are more likely to have
394 severe asthma that is insensitive to steroid therapy and are more commonly associated with

395 non-eosinophilic endotypes of disease (Hansbro et al., 2011; Wang et al., 2010; Wood et al.,
396 2010).

397 Several studies have highlighted potential roles for the NLRP3 inflammasome in the
398 development of allergic asthma. The inflammasome activator ATP has been shown to be
399 elevated in the airways of asthmatics compared to non-asthmatics, and is further increased
400 following challenge with allergen (Idzko et al., 2007; Muller et al., 2011). Experimental studies
401 using murine models of allergic airways disease (AAD) are being used to elucidate the role of
402 the inflammasome in the development of allergic asthma. These murine models typically
403 involve systemic sensitisation to model protein allergens (e.g. ovalbumin [Ova]) in the
404 presence of the Th2-inducing adjuvant alum, which is a known activator of the NLRP3
405 inflammasome (Hornung et al., 2008). A study by Eisenbarth *et al.*, used NLRP3-deficient mice
406 to demonstrate that this inflammasome is required for adjuvanticity of alum in allergic
407 antibody responses to antigen (Eisenbarth et al., 2008). Another study demonstrated the
408 importance of NLRP3 in allergic airway inflammation using an adjuvant (alum)-free Ova
409 model. Besnard *et al.*, used mice deficient in NLRP3, IL-1 receptor (IL-1R)1, IL-1 β or IL-1 α to
410 demonstrate critical roles for NLRP3-mediated IL-1 β responses in Ova-induced allergic airway
411 inflammation. They also reported that each of these factor-deficient mice exhibited marked
412 decreases in the production of Ova-induced, Th2-associated cytokines (Besnard et al., 2011).
413 Primiano *et al.*, further implicated the NLRP3 inflammasome in driving AAD, by showing that
414 therapy with the NLRP3 specific inhibitor, MCC950 reversed neutrophilic inflammation in AAD
415 (Primiano et al., 2016). In contrast, a different study by Kool *et al.*, demonstrated that uric
416 acid potently induces Th2 cell immunity in an NLRP3-independent, PI3K δ -dependent manner
417 (Kool et al., 2011). These findings are supported by a study by Allen *et al.*, which showed that
418 WT and NLRP3-deficient mice exhibited no differences in the key features of acute or chronic
419 Ova-induced AAD including eosinophilic airway inflammation, mucus hypersecretion and
420 airways hyperresponsiveness (AHR) (Allen et al., 2012). Similar observations have recently
421 been found with a combined particulate matter/Ova-induced model (Hirota et al., 2015).
422 Taken together, the role of the NLRP3 inflammasome in the pathogenesis of allergic asthma
423 remains to be elucidated.

424 It is likely that additional challenges, such as particulate matter or infections are needed
425 to drive inflammasome activation and more severe disease (Hansbro et al., 2011; Hansbro et
426 al., 2013). Recent clinical studies have identified that moderate to severe asthmatics have

427 increased Th1 and/or Th17 type, monocyte- or neutrophil-dominated immune responses in
428 their airway secretions (Baines et al., 2011; Simpson et al., 2007; Simpson et al., 2014a). This
429 has led to an increased focus on the development of more targeted therapies, particularly as
430 monocyte/neutrophil dominated asthma is more likely to be resistant to mainstay anti-
431 inflammatory corticosteroid therapy (Hansbro et al., 2011; Hansbro et al., 2013).

432 Recently, there has been an intense focus on elucidating the mechanisms of SSI asthma
433 and an increasing number of clinical and experimental studies strongly, and specifically,
434 implicate NLRP3 inflammasome activation and/or excess IL-1 β production in the pathogenesis
435 of this disease (Baines et al., 2011; Besnard et al., 2012; Essilfie et al., 2011; Hastie et al., 2010;
436 Kim et al., 2014a; Kim et al.; Kim et al., 2016; Kim et al., 2014b; Konno et al., 1996; Simpson
437 et al., 2014a; Starkey et al., 2013a; Starkey et al., 2014). Baines *et al.*, used gene expression
438 profiling of induced sputum to identify distinct gene signatures in different asthmatic
439 inflammatory endotypes. The expression of genes associated with the IL-1 β signalling
440 pathway, such as IL-1 β , IRAK2, IRAK3, IL-1R2, were significantly increased in the sputum of
441 neutrophilic asthmatics, which are more likely to be associated with severe asthma (Baines et
442 al., 2011). More recently, Simpson *et al.*, showed that neutrophilic asthmatics have increased
443 levels of NLRP3, Casp1 and IL-1 β expression in the airways, and macrophages and neutrophils
444 were the dominant cellular sources of NLRP3 and Casp1 in this cohort (Simpson et al., 2014a).
445 These asthmatics also had elevated expression of TLR2, TLR4, and IL-8 and increased levels of
446 LPS suggesting that innate immune activation in asthma may drive aberrant inflammasome
447 activation (Simpson et al., 2014a).

448 Substantial clinical evidence links bacterial respiratory infections with SSI asthma.
449 *Chlamydia pneumoniae* is an obligate intracellular bacterial pathogen that is associated with
450 SSI asthma. *Chlamydia*-associated asthma is less responsive to steroid treatment, and acute
451 antibody responses to *Chlamydia* strongly predicted the presence of neutrophils in sputum in
452 these patients (Cho et al., 2005; Patel et al., 2010; Wark et al., 2002). Airway neutrophilia also
453 positively predicted the presence of *Chlamydia* infection in SSI asthma (Patel et al., 2010).
454 Studies of *Chlamydia* are difficult due to the complexity of detection and the need to sample
455 the lower respiratory tract, where it prefers to grow. This may explain some negative studies.
456 *H. influenzae* is a Gram-negative bacteria that is the most commonly isolated bacterium from
457 the airways of SSI asthmatics (Simpson et al., 2007; Wood et al., 2010) compared to mild to
458 moderate asthmatics. We have developed and used experimental models of SSIAAD to assess

459 the effect of *Chlamydia* and *H. influenzae* respiratory infection on the development of SSIAAD,
460 to better understand their potential role in SSI asthma. We have shown that both infections
461 induce neutrophilia, and Th1 and/or Th17 responses and SSIAAD (Essilfie et al., 2015; Essilfie
462 et al., 2012; Essilfie et al., 2011; Horvat et al., 2010a; Horvat et al., 2010b; Kim et al.).
463 Significantly, both *Chlamydia* and *Haemophilus* respiratory infections induce the release of
464 active IL-1 β in an NLRP3 inflammasome-dependent, Casp1-mediated manner (Essilfie et al.,
465 2015; Essilfie et al., 2011; He et al., 2010; Horvat et al., 2010b; Kim et al.; Rotta Detto Loria et
466 al., 2013). Collectively, these data suggest that infection-induced, inflammasome-mediated
467 IL-1 β responses may play a key role in the development of SSI asthma.

468 Emerging evidence shows that systemic inflammation (determined through increased
469 serum levels of IL-6 and TNF- α) as a result of high-fat diet and/or obesity is associated with
470 severe asthma. Obese asthmatics are more likely to have severe disease and SSI asthma
471 (Forno et al., 2011; Gibeon et al., 2013; Scott et al., 2016). A recent study by Scott *et al.*,
472 showed that obese asthmatics had increased levels of systemic IL-6 and C-reactive protein,
473 and that these factors were positive predictors of neutrophilia in female obese asthmatics,
474 compared to non-obese female asthmatics (Scott et al., 2016). Importantly, IL-1 β levels are
475 significantly increased in the plasma of overweight (BMI of 25-29.9) and obese (BMI \geq 30)
476 women, compared to females in the normal weight range (Um et al., 2004), and obese
477 asthmatics, particularly obese females, are more likely to have severe asthma (Forno et al.,
478 2011; Gibeon et al., 2013; Lefaudeux et al., 2016; Scott et al., 2016). Recently, Kim *et al.*,
479 assessed the mechanisms of obesity-induced AHR in an experimental murine model of high
480 fat diet-induced obesity (Kim et al., 2014b). They showed that obese mice had increased AHR
481 in the absence of allergic sensitisation compared to non-obese control mice. They also
482 demonstrated that AHR in this model was driven by aberrant NLRP3 inflammasome-
483 dependent responses in the adipose tissue, which contributed to an induction of innate
484 lymphoid cells and increased IL-17 responses in the lung that led to spontaneous AHR (Kim et
485 al., 2014b). These experimental data highlight a key mechanism by which this inflammasome
486 may contribute to AHR in severe asthma, in the absence of allergic disease. Everaere *et al.*,
487 recently extended these findings by showing that obese mice with house dust mite (HDM)-
488 induced AAD had worsened AHR compared to non-obese, HDM allergen-challenged mice
489 (Everaere et al., 2016). Nevertheless, the roles of inflammasomes in obese AAD are yet to be
490 fully defined.

491 Further investigations and specific targeting of inflammasomes in the airways of allergen
492 challenged mice, particularly in the context of SSI asthma, is required to improve the
493 understanding of how NLRP3 inflammasomes contribute to the development of severe
494 asthma.

495

496 **6.3 Inflammasomes in COPD**

497 COPD is a progressive, obstructive disease of the lungs that encompasses several
498 conditions, including chronic bronchitis and emphysema, it is now the third leading cause of
499 death worldwide and its prevalence is increasing (Chapman et al., 2006; Fricker et al., 2014;
500 Keely et al., 2012; Lozano et al.). Significantly, several clinical studies have shown that IL-1 β
501 levels are elevated in the lungs of patients with COPD and that these levels increase further
502 during exacerbations of disease (Aaron et al., 2001; Chung, 2001). Clinical studies have also
503 shown that cigarette smoke induces the release of IL-1 β in the lung (Kuschner et al., 1996;
504 Pauwels et al., 2011). This is supported by data from mouse models that have shown
505 increased lung IL-1 β expression during cigarette smoke-induced experimental COPD (Beckett
506 et al., 2013). Pauwels *et al.*, interrogated a murine model of cigarette smoke-induced
507 pulmonary inflammation and showed that airway inflammation was significantly attenuated
508 through the neutralisation of IL-1 β (Pauwels et al., 2011). Cigarette smoke contains over 4,000
509 toxins, including LPS, which are capable of triggering innate immune responses through PRR
510 activation. These data therefore suggest that aberrant inflammasome activation may play an
511 important role in the pathogenesis of COPD.

512 Pouwels *et al.* showed that cigarette smoke exposure induces necroptosis, a form of
513 programmed necrosis, of airway epithelial BEAS-2B cells *in vitro* that results in the release of
514 endogenous DAMPs (Pouwels et al., 2016). Indeed, chromatin-binding high motility group box
515 1 protein (HMGB1) is a DAMP that can occur at high levels in the airways. HMGB1 levels are
516 elevated in the sputum and bronchoalveolar lavage fluid (BALF) of patients with COPD
517 (Ferhani et al., 2010; Hou et al., 2011). Significantly, HMGB1 has been shown to activate the
518 NLRP3 inflammasome in a TLR4-dependent manner in a model of haemorrhagic shock
519 syndrome (Xiang et al., 2011) suggesting that cigarette smoke-induced, HMGB1-mediated
520 activation of the inflammasome may play an important role in the pathogenesis of COPD.

521 Further mechanistic studies by Franklin *et al.*, showed that inflammasome responses may
522 also be mediated by the accumulation of ASC specks in the lungs of patients with COPD

523 (Franklin et al., 2014). ASC is an essential component of the NLRP3 and AIM2 inflammasomes
524 and is essential for Casp1 recruitment. Chronic activation of the inflammasome that results in
525 pyroptosis leads to the release of ASC specks, which have prion-like activity. They accumulate
526 in extracellular spaces and retain their ability to mature IL-1 β in the extracellular environment
527 (Franklin et al., 2014). These ASC specks are then readily phagocytosed by macrophages and
528 induce the production of IL-1 β in these cells. Most importantly, ASC specks are upregulated
529 in BALF from COPD patients and murine models of cigarette smoke-induced COPD (Franklin
530 et al., 2014). These studies indicate that ASC specks may drive aberrant inflammasome
531 responses and play an important role in the pathogenesis of COPD. In contrast, Di Stefano *et*
532 *al.*, found no correlation between NLRP3, Casp1 and IL-1 β responses in a cohort of stable
533 COPD patients compared to healthy smokers in a randomised control trial although they
534 proposed that they would be relevant in exacerbations (Di Stefano et al., 2014). Collectively,
535 these data highlight that the inflammasome may not be playing a role in stable COPD,
536 however, these studies do not assess the precise role of the inflammasome during the
537 development, progression or exacerbation of disease, which may be the critical issue. Thus,
538 the role of the NLRP3 inflammasome in COPD is complex and warrants further investigation
539 to delineate its roles. The use of mouse models that accurately replicate the major hallmark
540 features of cigarette smoke-induced COPD and exacerbations in a reasonable time frame and
541 in parallel with complementary human studies will be valuable in elucidating the mechanisms
542 involved, identifying new therapeutic targets and testing new therapies in this and other
543 respiratory diseases (Beckett et al., 2013; Conickx et al., 2016; Franklin et al., 2014; Fricker et
544 al., 2014; Hansbro et al., 2014; Haw et al., 2016; Hsu et al., 2015; Jarnicki et al., 2016; Liu et
545 al.; Simpson et al., 2014b; Starkey et al., 2013b; Tang et al., 2016; Tay et al., 2015) (Franklin et
546 al., 2014). Although not discussed here the role of the lung and gut microbiomes may also
547 play significant roles in inflammasome activity in these diseases, which could be elucidated
548 using similar strategies (Budden et al., 2016; Chambers et al., 2014; Ormerod et al., 2016).

549

550 **6.4 Inflammasomes in other chronic airway diseases**

551 **6.4.1 Pulmonary fibrosis**

552 Pulmonary fibrosis refers to a range of lung disorders characterised by irreversible
553 destruction and remodeling of lung architecture that occurs as a result of excess deposition
554 of collagen and extracellular matrix proteins. This results in scarring of the airways, which

555 leads to the significant breathing difficulties that are characteristic of the disease (dos Santos
556 et al., 2012). The role of the inflammasome in the pathogenesis of pulmonary fibrosis is
557 unclear, however, it is known that fibrosis-inducing irritants injure the lung epithelium (e.g.
558 silica, asbestos, cigarette smoke and bleomycin), and these are also known to directly activate
559 the NLRP3 inflammasome (Dostert et al., 2008; Gasse et al., 2007; Hornung et al., 2008). IL-
560 1 β secretion also promotes the production of TGF- β 1, a potent pro-fibrotic cytokine (Liu,
561 2008), and promotes neutrophil chemoattraction, which may contribute to epithelial
562 damage. However, the precise mechanisms of inflammasome-mediated pathogenesis in
563 pulmonary fibrosis are yet to be elucidated.

564

565 **6.4.2 Cystic fibrosis**

566 Cystic fibrosis (CF) is a debilitating lung disease that is caused by a genetic mutation in
567 the gene encoding the CF transmembrane conductance regulator (CFTR) (Caplen et al., 1995).
568 CFTR is a chloride ion transport channel that is defective in patients with CF. This results in
569 salt imbalances and excess accumulation of mucus in the lung that significantly increases the
570 risk of infections. The role of the inflammasome is not well understood in this disease,
571 however, the CFTR gene has been implicated as an important regulator of IL-1 β release
572 (Reiniger et al., 2007). A recent study by Iannitti *et al.*, used *in vitro* and *in vivo* models to
573 assess the importance of the NLRP3 and NLRC4 inflammasomes in the clearance of infections
574 in CF (Iannitti et al., 2016). However, they show that the deleterious effects of inflammation
575 caused by inflammasome activation are caused by NLRP3, which correlates with defective
576 NLRC4-IL-1R1 responses (Iannitti et al., 2016). These data distinguish important differences in
577 the roles of the NLRP3 and NLRC4 inflammasome in CF. They highlight that the activation of
578 different inflammasomes can contribute to inflammation in different ways, highlighting some
579 uniqueness in inflammasome-specific immune responses.

580

581 **6.5.2 Acute respiratory distress syndrome**

582 Acute respiratory distress syndrome (ARDS) refers to acute lung injury (ALI) in its most
583 severe form. ARDS can occur as a secondary complication of a range of different disorders
584 such as sepsis, ischemia, and trauma. It is a severe disease and these patients have a survival
585 rate of ~25% (Spragg et al., 2010). IL-18 is significantly increased in the blood of ARDS patients,
586 highlighting a potential role of inflammasome activation in this disease. Murine models of

587 LPS-induced ALI, which model ARDS, have determined that extracellular ATP is an important
588 neutrophil chemoattractant in the late phase of injury, and specifically targeting this factor
589 could limit damage induced by these cells. It is likely that the events observed in this model
590 are driven by aberrant NLRP3 inflammasome activation (Shah et al., 2014), where
591 extracellular ATP activates the NLRP3 inflammasome via the P2X₇R. Significantly, Wang *et al.*,
592 recently showed that pharmacological inhibition of P2X₇R suppresses the production IL-1 β
593 and Casp1, and ameliorates the key features of experimental ALI (Wang et al., 2015). These
594 data highlight the importance of the NLRP3 inflammasome in this form of chronic lung disease
595 and further support therapeutically targeting it.

596

597 **7. Conclusions**

598 Inflammasomes play a critical role in early innate immune responses particularly during
599 the resolution of infections in the lung. However, excessive inflammasome activation has
600 been associated with several major chronic inflammatory conditions. Whilst aberrant NLRP3
601 inflammasome responses are associated with SSI forms of asthma, COPD studies have not
602 clarified the causal nature of the relationship, let alone interrogated the potential for
603 therapeutic targeting of the NLRP3 inflammasome. Much of this is due to the lack of
604 understanding of upstream drivers of inflammasome assembly and activation, and use of
605 representative animal and experimental models that recapitulate the hallmark features of
606 disease. These are crucial in understanding the molecular mechanisms of action of the NLRP3
607 inflammasome in chronic airway disease and developing and testing new therapies.
608 Furthermore, it will be important to assess the contribution of other inflammasomes, such as
609 the AIM2 inflammasome, in the pathogenesis and exacerbation of these diseases. This is a
610 nascent field of enquiry that requires further investigation in order to elucidate the relative
611 contributions of the different inflammasomes and other IL-1 β -activating mechanisms in the
612 pathogenesis of disease. Current therapeutic strategies that globally target IL-1 β , such as the
613 Canakinumab, Anakinra and Rilanoccept biologics, rather than target excess production in a
614 pathway-specific way, may predispose to increased infection and are only delivered
615 systemically, rather than tissue-specifically, which will increase off-target effects. For the
616 treatment of excess IL-1 β responses, the development of inflammasome-mediated, site-
617 specific therapeutics may be more beneficial in suppressing inflammasome-associated
618 disease, whilst not predisposing to infection. To achieve this we need a greater understanding

619 of the molecular mechanisms driving inflammasome-associated disease, which may be
620 informed through the development and use of representative *in vivo* models and
621 complementary human studies.

622

623 8. References

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1119 **Table 1.** Exogenous activators of the inflammasome in the lung

Inflammasome	Activated by	Lung disease	Reference
NLRP1	Anthrax lethal toxin	Anthrax causing pneumoniae and severe respiratory collapse	(Banks et al., 2006)
NLRP3	<i>S. pneumoniae</i>	Resolution of Community acquired pneumonia (CAP)	(Mariathasan and Monack, 2007)
	<i>K. pneumoniae</i>		
	<i>Chlamydia spp.</i>	Severe, steroid-insensitive (SSI) asthma	(Essilfie et al., 2015; He et al., 2010; Rotta Detto Loria et al., 2013)
	<i>H. influenzae</i>		
	Influenza A virus		
	Asbestos	Asbestosis	(Dostert et al., 2008)
	Silica		(Hornung et al., 2008)
	Particulate matter (PM ₁₀)	Asthma	(Hirota et al., 2015; Hirota et al., 2012)
	Cigarette smoke		(Yang et al., 2015)
Lipopolysaccharide (LPS)	Chronic obstructive pulmonary disorder (COPD)	(Wang et al., 2015)	
	Acute lung injury (ALI)		
AIM2	<i>F. tularensis</i>	Resolution of CAP	(Fernandes-Alnemri et al., 2010)

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1122 **Fig 1.** Proposed model for the triggering of inflammasomes that leads to the development of severe,
 1123 steroid-insensitive (SSI) asthma and potentially other respiratory diseases. Infection and/or exposure
 1124 to infections, allergens, cigarette smoke, airway pollutants or other noxious stimuli in the asthmatic
 1125 airway triggers the assembly and activation of the NLRP3 inflammasome in the lung. This results in the
 1126 cleavage of pro-IL-1 β into active IL-1 β leading to increases in Th1- and/or Th17-associated responses
 1127 and neutrophils in the airways. This contributes to the development of inflammation, mucus
 1128 hypersecretion and airways hyperresponsiveness (AHR), which are resistant to steroid therapy. In the

1129 obese asthmatic lung, obese adipose tissue contributes to the activation of the NLRP3 inflammasome
1130 systemically. This results in an increase in systemic inflammation which activates the NLRP3
1131 inflammasome in the lung, resulting in steroid-insensitive asthma.