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A methanol and protic ionic liquid Ugi multicomponent reaction path to cytotoxic α -phenylacetamido amides†

 Ahmed Al Otaibi,^{‡a} Fiona M. Deane,^a Cecilia C. Russell,^a Lacey Hizartzidis,^a Siobhann N. McCluskey,^a Jennette A. Sakoff^b and Adam McCluskey^{*,a}

The Ugi four component reaction of an aldehyde, amine, isocyanide and an ethanoic acid was effected smoothly in protic ionic liquids ethylammonium nitrate (EAN) and propylammonium nitrate (PAN) to afford analogues of α -phenylacetamido amides in good to excellent isolated yields. The corresponding reactions in [BMIM][PF₆] and the protic ionic liquid ethanolammonium nitrate (ETAN) failed. Microwave irradiation in EAN facilitated rapid access to three focused libraries, based on the parent isocyanide: cyclohexyl isocyanide, benzyl isocyanide and ethyl isocyanoacetate. Analysis of the structure activity relationship data suggested the presence of a bulky moiety originating from the isocyanide (cyclohexyl and benzyl) enhanced cytotoxicity. Removal of the acetylenic H-atom from the ethanoic acid moiety was detrimental to cytotoxicity. The most active analogues produced, *N*-(2-cyclohexylamino)-1-(4-methoxyphenyl)-2-oxoethyl-*N*-(3,5-dimethoxyphenyl) propiolamide, returned average GI₅₀ values of ≤ 1 μ M across the cancer cell lines evaluated. Combined, these data suggest that analogues of this nature are interesting potential anti-cancer development leads.

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Introduction

The ability to rapidly probe the subtle nuances of chemical space through the decoration of simple and complex chemical scaffolds is critical to the development of new therapeutics and new therapeutic lead compounds.^{1–7} Medicinal chemistry relies on a robust toolkit of chemical modification broadly applicable across a diverse array of chemotypes. These robust, reliable synthetic tools enable rapid development of structure activity relationship data, the installation of favourable physicochemical properties and the bioisosteric replacement of toxicophores that may stymy drug development endeavours.^{8–10}

Multicomponent reactions (MCRs) are key ‘medicinal toolkit’ reactions permitting rapid assembly of three or more components in a one-pot sequence to produce a final product.^{11–20} MCRs present the medicinal chemist with additional benefits beyond rapid compound access and diversity, they are typically highly atom economic producing less waste

and require fewer resources, less time, energy and human effort. In many cases simpler purification processes and high convergence are also evident.^{13,21–24} Isocyanide-based MCRs are one of the most important and widely used of all the MCRs, and have been extensively used in the development of biologically active compounds.^{4,13,24–28}

The Ugi reaction is a well explored MCR for the generation of peptide like adducts,^{29–33} and as others and we have previously reported for the synthesis of heterocycles, *e.g.* quinolin-2-(1*H*)-ones and tetrahydroepoxisindole carboxamides.^{34,35} In these programs we have an on-going interest in the development of cytotoxic compounds, as have others. Previously Yamada *et al.* demonstrated the 4-component Ugi reaction provided robust access to a small library of cytotoxic propynoic acid carbamoyl methyl amide (PCMA) derivatives such as **1** and **2** (Fig. 1).³⁶

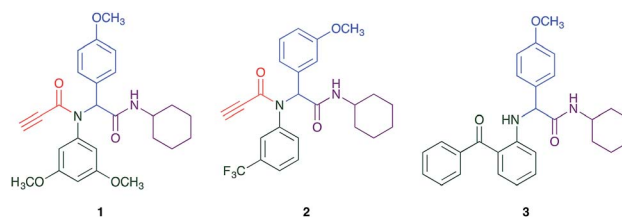


Fig. 1 Two examples for cytotoxic propynoic acid carbamoyl methyl amides (PCMA) synthesised by Yamada,³⁶ and 3-component Ugi product from Gordon *et al.*³⁴

^aChemistry, School of Environmental & Life Sciences, The University of Newcastle, University Drive, Callaghan, NSW 2308, Australia. E-mail: Adam.McCluskey@newcastle.edu.au

^bDepartment of Medical Oncology, Calvary Mater Newcastle Hospital, Edith Street Waratah 2298, Australia

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‡ Present address: Department of Chemistry, Faculty of Science, University of Hail, Hail, Kingdom of Saudi Arabia.



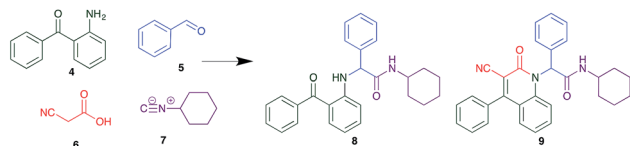
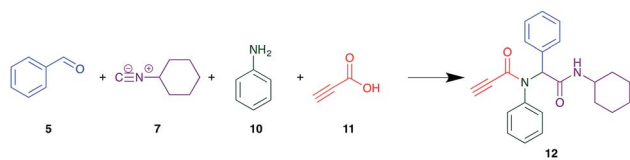
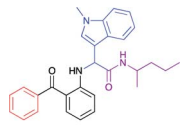
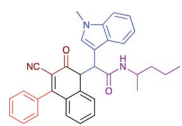
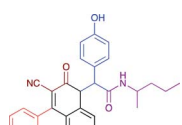
Scheme 1 Reagents and conditions: CH₃OH, rt.

Table 1 Effect of microwave irradiation time and reaction temperature on the isolated yield of the Ugi coupling of 5, 7, 10 and 11 in methanol to yield 12

Entry	Time (min)	Temp. °C	Yield%
1	5	50	11
2	5	100	58
3	5	120	63
4	10	120	69
5	20	120	83
6	30	120	78
7	1 h	120	70

Scheme 2 Reagents and conditions: CH₃OH, rt – 120 °C.Table 2 Evaluation of the cytotoxicity (GI₅₀ (μM)) of α-amino amide analogue (13) and quinolin-2-(1H)-one analogues (14 and 15) against a panel of ten cancer cell lines. GI₅₀ is the concentration of drug that reduces cell growth by 50% relative to an untreated control

Compound	HT29 ^a	SW480 ^a	MCF-7 ^b	A2780 ^c	H460 ^d	A431 ^e	Du145 ^f	BE2-C ^g	SJ-G2 ^h	MIA ⁱ
 13	6.4 ± 0.8	6.0 ± 0.4	6.0 ± 2.5	7.0 ± 1.5	12 ± 0	11 ± 0	11 ± 1	5.4 ± 0.9	6.7 ± 0.9	6.9 ± 1.2
 14	5.3 ± 0.3	11 ± 1	4.6 ± 1.1	3.9 ± 0.3	5.2 ± 0.1	2.7 ± 0.3	13 ± 0	3.6 ± 0.1	6.5 ± 0.3	6.0 ± 0.1
 15	8.7 ± 0.5	17 ± 1	7.9 ± 1.0	7.5 ± 0.6	11 ± 1	6.3 ± 0.5	18 ± 1	7.3 ± 0.3	10 ± 1	13 ± 1

^a HT29 and SW480 (colon carcinoma). ^b MCF-7 (breast carcinoma). ^c A2780 (ovarian carcinoma). ^d H460 (lung carcinoma). ^e A431 (skin carcinoma). ^f Du145 (prostate carcinoma). ^g BE2-C (neuroblastoma). ^h SJ-G2 and U87 (glioblastoma). ⁱ MIA (pancreatic carcinoma).

Table 3 The effect of PIL, microwave irradiation time and temperature on the yield of the Ugi coupling of 5, 7, 10 and 11 to yield 12

Entry	PIL	Irradiation time				
		50 °C	100 °C	120 °C	120 °C	120 °C
1	EAN	— ^a	58%	60%	62%	61%
2	PAN	— ^a	51%	55%	60%	54%
3	ETAN	— ^a	— ^a	— ^a	— ^a	— ^a

^a No reaction, starting material recovered.

Results and discussion

The PCMA compounds 1 and 2 in Fig. 1 are related to a series of Ugi-products that we previously reported differing primarily the incorporation of an aminoketone, *e.g.* 3,³⁴ but with these analogues we did not examine the cytotoxicity. In our hands a typical synthesis of this type of compound comprised the stirring of a methanolic solution of 2-aminobenzophenone (4) with benzaldehyde (5) followed by the addition of cyanoacetic acid (6) and cyclohexylisocyanide (7) afforded access to a separable mixture of the Ugi product, 8 and the Ugi-Knoevenagel product 9, Scheme 1. However, each reaction required 48 h to afford a modest 55% isolated yield.³⁴

As one of our team's drivers is enhancing green approaches to biologically active compounds, we explored alternative approaches that have been shown in other instances to

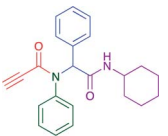
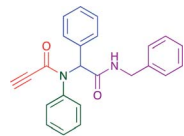
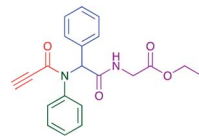
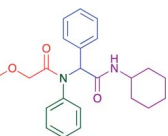
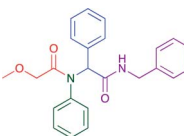
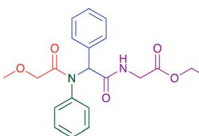
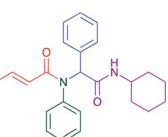
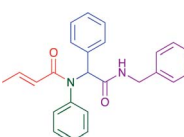
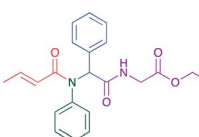
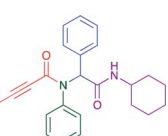
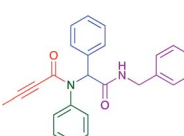
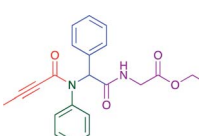
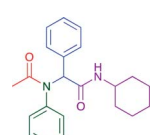
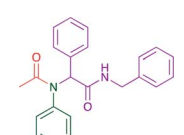
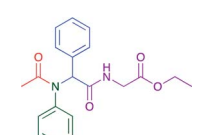
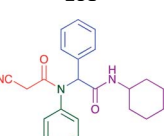
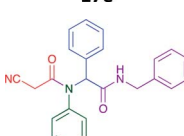
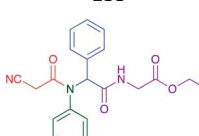
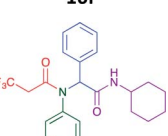
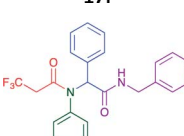
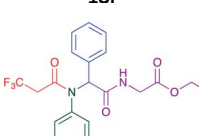


enhance, and accelerate, reaction efficiency.^{37–44} Microwave irradiation of a model system, only expected to produce the linear Ugi product, comprising 5, 7, 10 and 11 in methanol was evaluated at 50 °C, 100 °C and 120 °C and the outcomes presented in Table 1 (Scheme 2).⁴⁵ After 5 min irradiation at 50 °C a precipitate was observed, however TLC analysis indicated a mixture of product and starting materials. Increasing the reaction temperature to 100 °C and then 120 °C for 5 min saw a significant increase in product yield (58% and 63% respectively), and in these instances the collected precipitate was pure

by ¹H NMR analysis (ESI[†]). Irradiation of the reaction mixture for 20 min at 120 °C gave the desired 12 in an 83% isolated yield, representing a considerable enhancement over the initial 55% yield and 48 h reaction time. Prolonged heating resulted in a lower isolated yield, presumably a function of product degradation.

Expansion of the range of amines, acids, aldehydes and isonitriles afforded access to a broader range of Ugi products (Scheme 1; Table 2 and ESI[†] for detail). Subsequent MTT phenotypic screening against a panel of ten cancer cell lines

Table 4 Isolated yields of α -amino amides from the Ugi reaction in EAN under microwave irradiation conditions^a

Product	Ethanol ^b (%)	EAN ^c (%)	Product	Ethanol ^b (%)	EAN ^c (%)	Product	Ethanol ^b (%)	EAN ^c (%)
	83	58		70	54		71	60
12			17a			18a		
	76	66		67	69		75	64
16b			17b			18b		
	65	49		71	51		68	51
16c			17c			18c		
	78	56		67	55		66	54
16d			17d			18d		
	73	51		70	41		66	69
16e			17e			18e		
	60	45		79	61		58	59
16f			17f			18f		
	45	36		41	30		50	45
16g			17g			18g		

^a Starting material recovered. ^b At 120 °C for 20 min and using MeOH. ^c At 120 °C by using microwave and EAN for 10 min.





Table 5 Evaluation of the cytotoxicity (GI_{50} (μM)) of α -amino amide analogues **12**, **16d**, **16g**, **17a**, **17e**, **18a**, **1** and **19–21** against a panel of eleven cancer cell lines and one normal cell line. GI_{50} is the concentration of drug that reduces cell growth by 50% relative to an untreated control

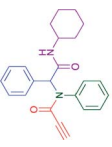
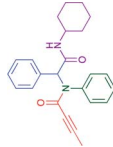
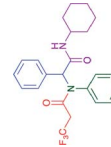
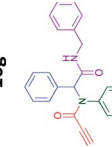
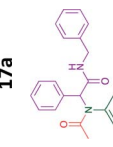
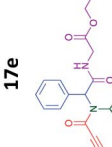
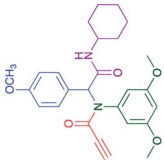
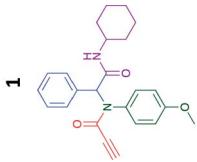
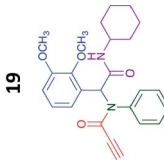
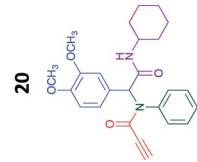
Compound	HT29 ^e	U87	MCF-7 ^b	A2780 ^c	H460 ^d	A431 ^c	Du145 ^f	BE2-C ^g	SJ-G2 ^h	MIA ⁱ	SMA ^j	MCF10A ^k
 12	0.27 ± 0.01	0.19 ± 0.01	1.6 ± 0.29	0.23 ± 0.02	3.6 ± 0.6	1.0 ± 0.2	0.18 ± 0.01	0.5 ± 0.2	3.0 ± 0.1	0.25 ± 0.02	0.25 ± 0.00	2.9 ± 0.1
 16d	15 ± 1	16 ± 0	4.6 ± 1.3	12 ± 0	13 ± 0.3	2.2 ± 0.1	8.6 ± 2.3	2.6 ± 0.3	13 ± 1	4.5 ± 0.3	14 ± 2	7.5 ± 1.8
 16g	5.9 ± 0.2	12 ± 1	8.0 ± 2.0	6.5 ± 0.2	6.0 ± 0.5	4.3 ± 0.3	4.8 ± 0.4	6.9 ± 0.6	11 ± 1	5.9 ± 0.1	10.0 ± 0.3	5.1 ± 0.5
 17a	0.17 ± 0.003	0.13 ± 0.017	1.7 ± 0.2	0.10 ± 0.034	2 ± 0.5	0.97 ± 0.2	1 ± 0.01	1.4 ± 0.3	2.5 ± 0.1	0.15 ± 0.009	0.16 ± 0.00	1.4 ± 0.4
 17e	> 50	> 50	> 50	16.0 ± 0.0	21 ± 0	13 ± 1	> 50	25 ± 9	15 ± 4	8.5 ± 0.9	> 50	15 ± 1
 18a	0.47 ± 0.09	0.27 ± 0.03	2.7 ± 0.13	0.33 ± 0.11	3.0 ± 0.4	2.1 ± 0.3	0.32 ± 0.04	1.8 ± 0.4	3.0 ± 0.2	0.50 ± 0.10	0.48 ± 0.06	6.6 ± 1.0



Table 5 (Contd.)

Compound	HT29 ^a	U87	MCF-7 ^b	A2780 ^c	H460 ^d	A431 ^e	Du145 ^f	BE2-C ^g	SJ-G2 ^h	MIA ⁱ	SMA ^j	MCF10A ^k
	0.07 ± 0.01	0.06 ± 0.02	1.70 ± 0.17	0.58 ± 0.51	1.4 ± 0.60	0.46 ± 0.15	0.11 ± 0.05	0.03 ± 0.01	1.70 ± 0.26	0.06 ± 0.03	ND	0.29 ± 0.03
1												
	0.20 ± 0.020	0.22 ± 0.00	1.2 ± 0.033	0.22 ± 0.017	2.2 ± 0.20	0.59 ± 0.26	0.20 ± 0.069	0.20 ± 0.0067	0.76 ± 0.57	0.20 ± 0.020	ND	3.3 ± 0.52
19												
	0.47 ± 0.12	0.71 ± 0.064	1.8 ± 0.067	0.49 ± 0.095	2.3 ± 0.30	1.2 ± 0.41	0.66 ± 0.24	0.46 ± 0.038	1.8 ± 0.088	0.46 ± 0.10	ND	7.6 ± 1.8
20												
	0.53 ± 0.041	0.73 ± 0.13	1.5 ± 0.12	0.54 ± 0.12	2.8 ± 1.1	1.0 ± 0.29	0.61 ± 0.25	0.27 ± 0.029	1.6 ± 0.18	0.44 ± 0.14	ND	5.1 ± 0.40
21												

^a HT29 and SW480 (colon carcinoma). ^b MCF-7 (breast carcinoma). ^c A2780 (ovarian carcinoma). ^d H460 (lung carcinoma). ^e A431 (skin carcinoma). ^f Du145 (prostate carcinoma). ^g BE2-C (neuroblastoma). ^h SJ-G2 and U87 (glioblastoma). ⁱ MIA (pancreatic carcinoma). ^j SMA (spontaneous murine astrocytoma). ^k MCF10A (normal breast). ND = not determined.

revealed modest levels of cytotoxicity at a single 25 μM compound concentration.⁴⁶ Only a small number of compounds, those with >80% growth inhibition at 25 μM against at least one cell line, from each of the α -amino amides (7) and quinolone (8) compound series being sufficiently active to proceed to full dose response evaluation (Table 2).

While the observed cytotoxicity of **13**, **14** and **15** was similar, with average GI_{50} values of 7.85, 6.18 and 10.7 μM respectively, access to the Ugi-Knoevenagel intramolecularly cyclised product series was limited by the nature of the acid (requiring the presence of an acidic methylene moiety). Accordingly, we limited further compound development and cytotoxicity screening to new families of α -amino amides.

Although microwave irradiation at 120 $^{\circ}\text{C}$ afforded excellent yields, this required the coupling to be conducted significantly above the boiling point of methanol with associated safety concerns. Accordingly, we sought a less volatile solvent, of which room temperature ionic liquids not only display the desired negligible to low vapour pressures, are environmentally friendly solvents, but as yet have not been explored for use in the Ugi coupling.⁴⁷ Thus, Ugi coupling of **5**, **7**, **10** and **11** was explored in $[\text{BMIM}][\text{PF}_6]$. After 24 h, despite allowing the imine from **3** and **5** to pre-form (Scheme 2), no product was observed at room temperature by TLC analysis. The same outcome was noted at 80 $^{\circ}\text{C}$ even after 24 h, but at 100 $^{\circ}\text{C}$ and 120 $^{\circ}\text{C}$, 20% and 32% product recovery was noted, however this was only after chromatographic separation.

In previous imine formation studies in our laboratory we noted an acceleration on imine formation on acid catalysis.^{35,48} We thus rationalised that this Ugi coupling might thus proceed more effectively in a protic ionic liquid.^{49,50} This is in keeping with prior reports of efficient multicomponent reactions in ionic liquids and deep eutectic salts.^{51–55} The model coupling described above in each of, ethylammonium nitrate (EAN), propylammonium nitrate (PAN) and ethanol ammonium nitrate (ETAN) with microwave irradiation was evaluated after 5 min irradiation at 50 $^{\circ}\text{C}$, 100 $^{\circ}\text{C}$ and 120 $^{\circ}\text{C}$ (Table 2). Pleasingly after 5 min irradiation at 100 $^{\circ}\text{C}$, with EAN, a 58% recovery of pure **12** was achieved, increasing to 60% at 120 $^{\circ}\text{C}$ with a similar outcome noted with PAN. Interestingly ETAN was ineffectual, presumably a consequence of the pendent OH moiety (Table 3). As EAN returned the highest yields of **12** at both 100 $^{\circ}\text{C}$ and 120 $^{\circ}\text{C}$, this PIL was used in subsequent experiments.

Having demonstrated the efficacy of the Ugi coupling in this model system, we next examined the production of three isocyanide derived compound libraries through the use of cyclohexyl isocyanide, benzyl isocyanide, and ethyl isocynoacetate, respectively, and a suite of diverse acids; propiolic, methoxyacetic, crotonic, 2-butynoic, acetic, cyanoacetic and 3,3,3-trifluoropropionic acids while retaining the parent aniline and benzaldehyde in the synthesis of three focused (**12** and **16b–g**; **17a–g**; and **18a–g**) libraries of α -amino amides (see Scheme 2). In most instances the yields in EAN were comparable to that obtained on microwave irradiation in MeOH at 120 $^{\circ}\text{C}$ (Table 4).

The cytotoxicity of the each of the focused libraries (**12** and **16b–g**, **17a–g** and **18a–g**) compounds, were investigated in

a modified panel of eleven tumour cell lines: colon (HT29, SW480), skin (A431), lung (H460), ovarian (A2780), breast (MCF-7), prostate (Du145), pancreatic (MIA), glioblastoma (SJ-G2, SMA, U87), and neuroblastoma (BE2-C) and one non-cancer derived normal breast cell line (MCF10A). Compounds were initially screened at 25 μM drug concentrations (ESI^{\dagger}), and from this screening, analogues **12**, **16d**, **16g**, **17a**, **17e** and **18a** showed sufficiently promising activity to warrant full dose evaluation, and these data are presented in Table 5.

Of the six analogues that proceeded to GI_{50} determination, exemplars from each isocyanide (cyclohexylisocyanide, benzylisocyanide and ethyl isocynoacetate) library were present. Four analogues contained an acetylene moiety (**12**, **16d**, **17a** and **18a**) with the other two derived from acetic (**17e**) and trifluoropropionic acid (**16g**). The cyclohexyl moiety was present in three of these analogues (**12**, **16d** and **16g**) and a benzyl moiety (**17a** and **17e**) in two, suggesting a requirement for a bulky hydrophobic moiety on the right-hand side of the molecule (as drawn). Analogues retaining the acetylenic moiety (**12**, **17a** and **18a**) were considerably more potent than the equivalent propargylic analogue (**16d**) with average GI_{50} values of 1.16 μM , 0.97 μM , 1.8 μM and 9.4 μM respectively. With the acetylenic series of compounds (**12**, **17a** and **18a**) sub-micro molar levels of toxicity is noted against HT29, U87, A2780, MIA and SMA cell lines. Removal of the acetylene (or propargylic) moiety with **17a** results in a >30 fold potency reduction (average GI_{50} value > 30 μM), but the introduction of a trifluoromethyl moiety with **16g** results in an analogue equipotent with **16d** with an average GI_{50} of 7.2 μM (Table 5).

Given the clear enhanced cytotoxicity of analogues containing the acetylenic moiety, and the similarity to those from the Yamada report,³⁰ we sought to directly compare both analogues sets and expand the pharmacophore of these compounds. Thus, we in-house synthesised lead **1** and three additional methoxy analogues (**19–21**) as per Scheme 2. Screening data for these four analogues (**1**, **19–21**) is presented in Table 5. Analysis of these data is consistent with the acetylene moiety analogues being sub-micromolar cytotoxic with all four analogues returning average GI_{50} values $\leq 1 \mu\text{M}$ for the cancer cell lines examined. Of note in this series of analogues was the removal of the aromatic methoxy moieties that saw a reduction in the observed toxicity towards the MCF10A normal breast cell line. This was most apparent on removal of those methoxy moieties originating from the starting aniline. Repositioning of the methoxy moieties to the starting benzaldehyde was tolerated in this series of analogues, with both the 2,3- and 3,4-dimethoxy disposed analogues showing a 10-fold selectivity for cancer the cancer cell lines *versus* the MCF10A normal cell line. This is a promising level of selectivity at this stage of compound exploration.

Conclusions

The Ugi coupling of an aldehyde, amine, isocyanide and ethanoic acid to afford analogues of α -phenylacetamido amides in modest yield under microwave irradiation conditions in methanol. Solvent switching to the protic ionic liquids EAN and PAN



afforded higher or comparable yields without the need to heat the reaction about the solvent boiling point as was the case with the methanol reaction, with reactions conducted at 120 °C. Neither [BMMIM][PF₆] nor ETAN afforded useful yields of the desired products. Our prior studies have shown that the initial imine formation requires acid catalysis, with the free OH moiety of ETAN presumed to interfere with this reaction. The Ugi coupling in the presence of an acid with an acidic methylene moiety afforded an additional Ugi-Knoevenagel cyclised product, but given the need for the acid component to contain an additional α -methylene requirement (which were of limited supply in our laboratory), our subsequent synthesis was directed at the four component Ugi products. From this, three focused libraries, based on the starting cyclohexylisocyanide, benzylisocyanide and ethyl isocyanacetate, were produced in good to excellent yield.

The acetylene containing **12**, **16d**, **17a** and **18a**; acetic acid **17e** and derived with the other two derived from acetic **17e** and trifluoropropionic acid derived **16g** displayed excellent broad spectrum cytotoxicity. Analysis of the structure activity relationship data suggested the presence of a bulky moiety originating from the isocyanide (cyclohexyl and benzyl) enhanced cytotoxicity. Removal of the acetylenic H-atom was detrimental to cytotoxicity. The acetylene H-atom retention was confirmed with the synthesis of Yamada's lead **1** and three additional analogues **19–21**,³⁰ which were the most active analogues produced with an average GI₅₀ of $\leq 1 \mu\text{M}$ across the cancer cell lines evaluated. Removal and repositioning of the *N*-(3,5-dimethoxyphenyl) moieties of **1** retained cancer cell line activity while reducing cytotoxicity against the MCF10A normal cell line. While these analogues display no greater cytotoxicity relative to those reported by Yamada,³⁶ they aid in the development of a PCMA cytotoxicity pharmacophore. Combined these data suggest that analogues **19–21**, in particular, are interesting potential anti-cancer development leads.

Experimental

Chemistry

General Methods. All reagents were purchased from Sigma-Aldrich, AK Scientific, Matrix Scientific or Lancaster Synthesis and were used without purification. All solvents were re-distilled from glass prior to use.

¹H and ¹³C NMR spectra were recorded on a Bruker Advance™ AMX 400 at 400.13 and 100.62 MHz, respectively and Advance™ AMX 600 at 600.21 and 150.92 MHz, respectively. Chemical shifts (δ) are reported in parts per million (ppm) measured relative to the internal standards. Coupling constants (*J*) are expressed in hertz (Hz). Mass spectra were recorded on a Shimadzu LCMS 2010 EV and Agilent 6100 series single quadrupole LCMS using a mobile phase of 1 : 1 acetonitrile : H₂O with 0.1% formic acid. The University of Wollongong, Australia, Mass Spectrometry User resource & Research Facility (MSURRF) analysed samples for High Resolution Mass Spectrometry HRMS Analytical HPLC traces were obtained using a Shimadzu system possessing a SIL-20A autosampler, dual LC-20AP pumps, CBM-20A bus module, CTO-

20A column heater, and a SPD-20A UV/vis detector. This system was fitted with an Alltima™ C₁₈ 5 μm 150 mm \times 4.6 mm column with solvent A: 0.06% trifluoroacetic acid (TFA) in water and solvent B: 0.06% TFA in CH₃CN–H₂O (90 : 10). In each case HPLC traces were acquired at a flow rate of 2.0 mL min⁻¹, gradient 10–100 (%B), over 15.0 min, with detection at 220 nm and 254 nm.

Melting points were recorded on a Büchi Melting Point M-565 instrument. IR spectra were recorded on a PerkinElmer Spectrum Two™ FTIR Spectrometer with the UATR accessories. Thin layer chromatography (TLC) was performed on Merck 60 F254 pre-coated aluminium plates with a thickness of 0.2 mm. Column chromatography was performed under 'flash' conditions on Merck silica gel 60 (230–400 mesh).

In vitro growth inhibition assays

All cell lines were cultured in a humidified atmosphere 5% CO₂ at 37 °C. The cancer cell lines were maintained in Dulbecco's modified Eagle's medium (DMEM) (Trace Biosciences, Australia) supplemented with 10% foetal bovine serum, 10 mM sodium bicarbonate penicillin (100 IU mL⁻¹), streptomycin (100 μg mL⁻¹), and glutamine (4 mM). The non-cancer MCF10A cell line was cultured in DMEM : F12 (1 : 1) cell culture media, 5% heat inactivated horse serum, supplemented with penicillin (50 IU mL⁻¹), streptomycin (50 μg mL⁻¹), 20 mM Hepes, *L*-glutamine (2 mM), epidermal growth factor (20 ng mL⁻¹), hydrocortisone (500 ng mL⁻¹), cholera toxin (100 ng mL⁻¹), and insulin (10 μg mL⁻¹). Cytotoxicity was determined by plating cells in duplicate in 100 mL medium at a density of 2500–4000 cells per well in 96 well plates. On day 0, (24 h after plating) when the cells were in logarithmic growth, 100 μL medium with or without the test agent was added to each well. After 72 h compound exposure growth inhibitory effects were evaluated using the MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) assay and absorbance read at 540 nm. An eight point dose response curve was produced from which the GI₅₀ value was calculated, representing the drug concentration at which cell growth was inhibited by 50% based on the difference between the optical density values on day 0 and those at the end of drug exposure.⁴⁰

Synthesis of α -amino amides

(*RS*)-*N*-(2-(Cyclohexylamino)-2-oxo-1-phenylethyl)-*N*-phenylpropionamide (**12**)

Method A (batch reaction and using MeOH). A solution of MeOH (5.0 mL), aniline (0.09 mL, 1.00 mmol), and benzaldehyde (0.10 mL, 1.00 mmol) was stirred at room temperature for 0.5 h. To the stirred solution was added propionic acid (0.06 mL, 1.00 mmol) followed by the addition of cyclohexyl isocyanide (0.11 mL, 1.00 mmol). The reaction mixture was stirred at room temperature for 24 h and the resulting precipitate was collected, washed with diethyl ether, and dried to afford (**12**) as a white solid without purification (0.20 g, 55%).

Method B (microwave reaction and using MeOH). A solution of MeOH (3.0 mL), aniline (0.09 mL, 1.00 mmol), and benzaldehyde (0.10 mL g, 1.00 mmol) was stirred at room temperature



for 5 min at room temperature. Then, propiolic acid (0.06 mL, 1.00 mmol) followed by the addition of cyclohexyl isocyanide (0.11 mL, 1.00 mmol) were added to the stirred solution. The microwave irradiation was applied for 20 min under 120 °C. After 20 minutes the formation of a precipitate was observed, the solid was collected and washed with diethyl ether and dried to obtain the desired compound as a white solid (0.30 g, 83%).

Method C (microwave reaction and using [EAN][HNO₃]). A solution of [EAN][HNO₃] (3.0 mL), aniline (0.09 mL, 1.00 mmol), and benzaldehyde (0.10 mL, 1.00 mmol) was stirred at room temperature for 5 min at room temperature. Then propiolic acid and cyclohexylcyanide were added sequentially to the stirred solution and the reaction mixture was then irradiated at 120 °C for 20 minutes. Following irradiation the reaction mixture was extracted with EtOAc, washed with H₂O and the combined organic layers dried over MgSO₄ and concentrated *in vacuo*. Finally, the crude reaction mixture was subjected to flash silica gel column chromatography (Hexane : EtOAc) to afford pure product (12) (0.21 g, 58%); LRMS (ESI⁺) *m/z* 361 [M + H]⁺; mp 194.1–196.7; IR (cm⁻¹): 3278, 2924, 2848, 2113, 1640, 1594, 1563, 1491, 1452, 1359, 1308, 1250, 1106, 762, 730, 649, 660, 569. ¹H NMR (400 MHz, CDCl₃): δ 7.10–7.25 (m, 10H), 6.01 (s, 1H), 5.58 (d, *J* = 8.2 Hz, 1H), 3.79–3.87 (m, 1H), 2.79 (s, 1H), 1.80–1.98 (m, 1H), 1.79–1.88 (m, 1H), 1.52–1.71 (m, 3H), 1.25–1.40 (m, 2H), 0.96–1.18 (m, 3H); ¹³C NMR (101 MHz, CDCl₃): δ 167.7, 153.9, 139.2, 133.9, 130.9, 130.5, 128.9, 128.6, 80.7, 76.2, 65.2, 49.0, 32.9, 25.6, 24.9.

(RS)-N-Cyclohexyl-2-(2-methoxy-N-phenylacetamido)-2-phenylacetamide (16b). Synthesised using the general procedure (Method b and c) as described for (12) by using aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), methoxyacetic acid (0.07 g, 1.00 mmol), and cyclohexyl isocyanide (0.11 mL, 1.00 mmol), to afford (16b) as a brown solid (Method b 76% and 66% c); LRMS (ESI⁺) *m/z* 381 [M + H]⁺; mp 210.1–214.3; IR (cm⁻¹): 3260, 3099, 2926, 2854, 1670, 1645, 1595, 1564, 1493, 1447, 1278, 1250, 1127, 976, 698, 557. ¹H NMR (400 MHz, CDCl₃): δ 7.07–7.30 (m, 10H), 6.05 (s, 1H), 5.58–5.63 (d, *J* = 8.0 Hz, 1H), 3.67–3.88 (m, 3H), 3.33 (s, 3H), 1.90–2.00 (m, 1H), 1.80–1.90 (d, *J* = 12.5 Hz, 1H), 1.24–1.41 (q, *J* = 11.6 Hz, 2H), 0.96–1.19 (m, 3H); ¹³C NMR (101 MHz, CDCl₃): δ 169.9, 168.5, 138.5, 134.4, 130.6, 130.6, 129.1, 128.6, 128.5, 71.0, 65.1, 59.4, 49.0, 33.0, 32.9, 25.6, 25.0, 24.9.

(RS)-(E)-N-(2-(Cyclohexylamino)-2-oxo-1-phenylethyl)-N-phenylbut-2-enamide (16c). Synthesised using the general procedure (Method b and c) as described for (12) above from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 mL, 1.00 mmol), crotonic acid (0.23 g, 1.00 mmol), and cyclohexyl isocyanide (0.11 mL, 1.00 mmol), to afford (16c) as a white solid (Method b 65% and c 49%); LRMS (ESI⁺) *m/z* 377 [M + H]⁺, mp 172.5–187.4; IR (cm⁻¹): 3266, 2926, 2848, 1666, 1645, 1627, 1447, 1367, 1237, 1107, 960, 750, 697, 558. ¹H NMR (400 MHz, CDCl₃): δ 7.02–7.24 (m, 10H), 6.90–7.02 (m, 1H), 6.03 (s, 1H), 5.74–5.84 (d, *J* = 8.0 Hz, 1H), 5.57–5.65 (dd, *J* = 15.0, 1.7 Hz, 1H), 3.77–3.89 (m, 1H), 1.90–1.99 (m, 1H), 1.81–1.90 (m, 1H), 1.52–1.73 (m, 5H), 1.24–1.41 (m, 3H), 0.97–1.20 (m, 3H); ¹³C NMR (101 MHz, CDCl₃): δ 168.7, 166.5, 142.3, 140.0, 134.9, 130.5, 130.2, 128.8, 128.3, 128.2, 127.9, 122.9, 65.6, 48.6, 32.9, 32.8, 25.5, 24.8, 24.7, 18.0.

(RS)-N-(2-(Cyclohexylamino)-2-oxo-1-phenylethyl)-N-phenylbut-2-enamide (16d). Synthesised using the general procedure (Method b and c) as described for (12) above from aniline (0.09 g, 1.00 mmol), benzaldehyde (0.10 mL, 1.00 mmol), 2-butynoic acid (0.23 g, 1.00 mmol), and cyclohexyl isocyanide (0.11 mL, 1.00 mmol) in MeOH (5.0 mL) to afford (16d) as a white solid (Method b 78% and c 56%); LRMS (ESI⁺) *m/z* 375 [M + H]⁺; mp 211.5–213.5; IR (cm⁻¹): 3266, 2925, 2854, 2256, 1649, 1633, 1553, 1493, 1358, 1249, 1101, 979, 697, 661, 570. ¹H NMR (400 MHz, CDCl₃): δ 7.12–7.23 (m, 10H), 5.97–6.03 (s, 1H), 5.63–5.72 (d, *J* = 8.2 Hz, 1H), 3.74–3.88 (m, 1H), 1.90–1.98 (dd, *J* = 12.8, 4.2 Hz, 1H), 1.80–1.89 (m, 1H), 1.65–1.70 (d, *J* = 7.3 Hz, 7H), 1.26–1.41 (m, 2H), 0.97–1.19 (m, 3H); ¹³C NMR (101 MHz, CDCl₃): δ 168.1, 155.2, 139.9, 134.3, 130.7, 130.3, 128.6, 128.5, 128.4, 128.2, 91.5, 74.1, 65.1, 48.9, 32.9, 32.9, 25.6, 24.9, 24.8, 4.0.

(RS)-N-Cyclohexyl-2-phenyl-2-(N-phenylacetamido)acetamide (16e). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), acetic acid (0.23 g, 1.00 mmol), and cyclohexyl isocyanide (0.11 mL, 1.00 mmol) to afford (16e) as a white solid (Method b 73%, and c 51%); LRMS (ESI⁺) *m/z* 351 [M + H]⁺; mp 186.1–189.4; 3260, 2926, 2848, 1645, 1563, 1493, 1382, 1250, 1240, 889, 730, 696, 647, 548, 515. ¹H NMR (400 MHz, CDCl₃): δ 7.06–7.25 (m, 10H), 5.99–6.03 (s, 1H), 5.56–5.64 (d, *J* = 7.8 Hz, 1H), 3.70–3.93 (m, 1H), 1.89–1.9945 (m, 1H), 1.84–1.88 (s, 3H), 1.75 (s, 1H), 1.52–1.71 (m, 3H), 1.25–1.41 (m, 2H), 0.96–1.18 (m, 3H); ¹³C NMR (101 MHz, CDCl₃): δ 171.3, 168.9, 140.9, 134.9, 130.4, 129.0, 128.4, 128.1, 65.2, 48.9, 33.0, 32.9, 25.6, 25.0, 24.9, 23.4.

(RS)-2-Cyano-N-(2-(cyclohexylamino)-2-oxo-1-phenylethyl)-N-phenylacetamide (16f). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), cyanoacetic acid (0.23 g, 1.00 mmol), and cyclohexyl isocyanide (0.11 mL, 1.00 mmol), to afford (16f) as a white solid (Method b 60% and c 45%); LRMS (ESI⁺) *m/z* 376 [M + H]⁺; mp 241.9–245.4; IR (cm⁻¹): 3253, 2928, 2858, 1668, 1643, 1594, 1563, 1493, 1386, 1239, 1104, 1005, 734, 697, 408. ¹H NMR (400 MHz, CDCl₃): δ 7.05–7.31 (m, 10H), 5.99–6.02 (s, 1H), 5.38–5.44 (d, *J* = 8.1 Hz, 1H), 3.75–3.87 (m, 1H), 3.1281–3.2897 (m, 2H), 1.89–1.99 (dd, *J* = 12.0, 4.1 Hz, 1H), 1.77–1.88 (m, 1H), 1.6245–1.71 (m, 2H), 1.23–1.42 (m, 3H), 0.93–1.17 (m, 3H); ¹³C NMR (101 MHz, CDCl₃): δ 167.9, 162.7, 138.6, 133.8, 130.5, 129.3, 129.1, 128.8, 65.8, 49.2, 33.0, 32.9, 26.4, 25.6, 25.0, 24.9.

(RS)-N-(2-(Cyclohexylamino)-2-oxo-1-phenylethyl)-3,3,3-trifluoro-N-phenylpropanamide (16g). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), 3,3,3-trifluoropropionic acid (0.23 g, 1.00 mmol), and cyclohexyl isocyanide (0.11 mL, 1.00 mmol) to afford (16g) as a white solid (Method b 45% and c 36%); LRMS (ESI⁺) *m/z* 377 [M + H]⁺; mp 202.9–211.9; IR (cm⁻¹): 3327 (NH), 3056 (CH), 2961, 2929 (CH); ¹H NMR (400 MHz, CDCl₃): δ 6.95–7.46 (m, 10H), 6.03 (s, 1H), 5.46–5.56 (d, *J* = 8.1 Hz, 1H), 3.74–3.86 (m, 1H), 2.83–3.02 (m, 2H), 1.89–1.98 (dd, *J* = 13.3, 4.3 Hz, 2H), 1.79–1.88 (m, 1H), 1.52–1.72 (m, 3H), 1.40–1.47 (d, *J* = 9.1 Hz,



1H), 1.24–1.40 (m, 2H), 0.93–1.17 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 168.2, 164.1, 164.1, 139.0, 134.1, 130.5, 128.9, 128.8, 128.6, 65.3, 49.0, 39.5, 39.2, 38.9, 38.6, 32.9, 32.9, 25.6, 24.1, 24.9.

(RS)-N-(2-(Benzylamino)-2-oxo-1-phenylethyl)-N-phenylpropiolamide (17a). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), propiolic acid (0.06 g, 1.00 mmol), and benzyl isocyanide (0.12 mL, 1.00 mmol), to afford (17a) as a white solid (Method b 70% and c 54%); LRMS (ESI⁺) *m/z* 369 [M + H]⁺, mp 184.1–194.9; IR (cm⁻¹): 3307 (NH), 3189, 1668, 1635 (C); ¹H NMR (400 MHz, CDCl₃): δ 7.12–7.32 (m, 15H), 6.03–6.05 (s, 1H), 5.97–6.03 (d, *J* = 6.1 Hz, 1H), 4.44–4.56 (m, 2H), 3.45–3.49 (s, 1H); ¹³C NMR (101 MHz, CDCl₃): δ 168.7, 153.9, 139.2, 137.9, 133.5, 130.9, 130.5, 128.8, 128.7, 128.7, 127.7, 127.6, 80.8, 65.4, 51.0, 44.0.

(RS)-N-Benzyl-2-(2-methoxy-N-phenylacetamido)-2-phenylacetamide (17b). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), methoxyacetic acid (0.07 mL, 1.00 mmol), and benzyl isocyanide (0.12 mL, 1.00 mmol) to afford (17b) as a white solid (Method b 67% and c 69%); LRMS (ESI⁺) *m/z* 389 [M + H]⁺. Mp 182.5–185.7; IR (cm⁻¹): 3255 (NH), 1664, 1651 (CO); ¹H NMR (400 MHz, CDCl₃): δ 7.07–7.33 (m, 15H), 6.62–6.68 (m, 1H), 6.09 (s, 1H), 4.46–4.52 (dd, *J* = 8.6, 5.8 Hz, 1H), 3.68–3.81 (d, *J* = 1.6 Hz, 2H), 3.42 (s, 1H), 3.34 (s, 2H); ¹³C NMR (101 MHz, CDCl₃): δ 170.0, 169.5, 138.4, 138.1, 133.9, 130.6, 130.5, 129.6, 129.2, 129.1, 128.8, 128.8, 128.7, 128.7, 128.6, 128.1, 127.7, 127.5, 119.7, 113.9, 71.0, 65.3, 59.4, 44.0.

(RS)-(*E*)-N-(2-(Benzylamino)-2-oxo-1-phenylethyl)-N-phenylbut-2-enamide (17c). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), crotonic acid (0.23 g, 1.00 mmol), and benzyl isocyanide (0.12 mL, 1.00 mmol), to afford (17c) as a white solid (Method b 71% and c 51%); LRMS (ESI⁺) *m/z* 385 [M + H]⁺; mp 208.9–221.7; IR (cm⁻¹): 3337 (NH), 1657, 1620 (CO); ¹H NMR (400 MHz, CDCl₃): δ 7.03–7.31 (m, 15H), 6.92–7.03 (m, 1H), 6.25 (s, 1H), 6.08 (s, 1H), 5.59–5.66 (dd, *J* = 15.1, 1.7 Hz, 1H), 4.48–4.54 (d, *J* = 5.8 Hz, 2H), 1.68–1.74 (dd, *J* = 7.0, 1.7 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃): δ 169.9, 166.7, 142.6, 140.1, 138.3, 134.7, 130.6, 130.4, 129.0, 128.7, 128.5, 128.5, 128.1, 127.7, 127.4, 123.0, 66.0, 43.9, 18.2.

(RS)-N-(2-(Benzylamino)-2-oxo-1-phenylethyl)-N-phenylbut-2-ynamide (17d). Synthesised using the general procedure (Method b and c) as described for (12) above from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), 2-butyric acid (0.23 g, 1.00 mmol), and benzyl isocyanide (0.12 mL, 1.00 mmol), to afford (17d) as a white solid (Method b 67% and c 55%); LRMS (ESI⁺) *m/z* 383 [M + H]⁺; mp 183.1–187.8; IR (cm⁻¹): 3298 (NH), 2249, 1164, 1629 (CO); ¹H NMR (400 MHz, CDCl₃): δ 7.26–7.30 (m, 2H), 7.16–7.25 (m, 13H), 6.07–6.12 (d, *J* = 6.1 Hz, 1H), 6.03 (s, 1H), 4.34–4.63 (m, 3H), 1.67 (s, 3H); ¹³C NMR (101 MHz, CDCl₃): δ 169.1, 155.2, 139.9, 138.0, 134.0, 130.7, 130.4, 128.8, 128.8, 128.6, 128.5, 128.3, 127.7, 127.5, 91.6, 74.1, 65.4, 44.0, 4.0.

(RS)-N-Benzyl-2-phenyl-2-(N-phenylacetamido)acetamide (17e). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), acetic acid (0.23 g, 1.00 mmol), and benzyl isocyanide (0.12 mL, 1.00 mmol), to afford (17e) as a white solid (Method b 70% and c 41%); LRMS (ESI⁺) *m/z* 359 [M + H]⁺; mp 185.2–188.7; IR (cm⁻¹): 3247 (NH), 1652, 1645 (CO); ¹H NMR (400 MHz, CDCl₃): δ 1.65–1.69 (s, 1H), 1.85–1.88 (s, 3H), 4.43–4.55 (m, 2H), 6.03–6.12 (d, *J* = 10.4 Hz, 2H), 7.06–7.32 (m, 15H); ¹³C NMR (101 MHz, CDCl₃): δ 171.4, 169.9, 140.9, 138.2, 134.6, 130.5, 130.4, 129.1, 128.8, 128.6, 128.5, 128.2, 127.7, 127.5, 65.4, 43.9, 23.4.

(RS)-N-Benzyl-2-(2-cyano-N-phenylacetamido)-2-phenylacetamide (17f). Synthesised utilising the general procedure described in (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), cyanoacetic acid (0.23 g, 1.00 mmol), and benzyl isocyanide (0.12 mL, 1.00 mmol), to afford (17f) as a brown solid (Method b 79% and c 61%); LRMS (ESI⁺) *m/z* 389 [M + H]⁺; mp 216.1–221.1; IR (cm⁻¹): 3256 (NH), 1664, 1654 (CO); ¹H NMR (400 MHz, CDCl₃): δ 3.15–3.29 (m, 2H), 4.41–4.55 (m, 2H), 5.91 (s, 1H), 6.05 (s, 1H), 7.08–7.29 (m, 15H); ¹³C NMR (101 MHz, CDCl₃): δ 168.9, 162.8, 138.6, 137.8, 130.6, 129.3, 129.2, 128.8, 128.8, 127.7, 127.7, 114.1, 65.9, 44.1, 26.5.

(RS)-N-(2-(Benzylamino)-2-oxo-1-phenylethyl)-3,3,3-trifluoro-N-phenylpropanamide (17g). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), 3,3,3-trifluoropropionic acid (0.23 g, 1.00 mmol), and benzyl isocyanide (0.12 mL, 1.00 mmol), to afford (17g) as a white solid (Method b 41% and c 30%); LRMS (ESI⁺) *m/z* 427 [M + H]⁺, mp 160.8–164.4; IR (cm⁻¹): 3266 (NH), 1666, 1654 (CO); ¹H NMR (400 MHz, CDCl₃): δ 7.54–7.79 (s, 1H), 7.14–7.31 (m, 11H), 7.09–7.13 (m, 2H), 6.42–6.68 (s, 1H), 6.06–6.10 (s, 1H), 5.93–6.00 (s, 1H), 4.38–4.58 (m, 2H), 2.82–3.06 (m, 2H); ¹³C NMR (101 MHz, CDCl₃): δ 169.0, 133.6, 130.4, 128.9, 128.7, 128.6, 127.6, 127.5, 95.0, 65.3, 43.9.

(RS)-Ethyl (2-phenyl-2-(N-phenylpropiolamido)acetyl)glycinate (18a). Synthesised using the general procedure (Method b and c) as described for (12) above from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), propiolic acid (0.06 g, 1.00 mmol), and ethyl isocyanoacetate (0.11 mL, 1.00 mmol), to afford (18a) as a white solid (Method b 71% and c 60%); LRMS (ESI⁺) *m/z* 365 [M + H]⁺; mp 170.5–172.2; IR (cm⁻¹): 3333 (NH), 1625 (CO); ¹H NMR (400 MHz, CDCl₃): δ 7.12–7.25 (m, 10H), 6.31–6.36 (s, 1H), 6.11–6.15 (s, 1H), 4.13–4.20 (q, *J* = 7.1 Hz, 2H), 4.02–4.06 (t, *J* = 5.1 Hz, 2H), 2.77–2.82 (s, 1H), 2.01–2.03 (s, 1H), 1.22–1.26 (t, *J* = 7.1 Hz, 4H). ¹³C NMR (101 MHz, CDCl₃): δ 169.6, 168.9, 153.9, 139.0, 133.2, 130.8, 130.6, 129.0, 128.7, 128.7, 128.6, 80.8, 76.1, 64.9, 61.6, 60.5, 41.8, 21.1, 14.2.

(RS)-Ethyl (2-(2-methoxy-N-phenylacetamido)-2-phenylacetyl)glycinate (18b). Synthesised using the general procedure (Method B and C) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 mL, 1.00 mmol), methoxyacetic acid (0.08 mL, 1.00 mmol), and ethyl isocyanoacetate (0.12 mL, 1.00 mmol), to afford (18b) as a white solid (Method B 0.29, 75% and Method C 0.25 g, 65%); LRMS (ESI⁺) *m/z* 385 [M + H]⁺; mp



170–178 °C; IR (cm⁻¹): 3330 (NH), 1627 (CO); ¹H NMR (400 MHz, CDCl₃) δ 7.26–7.13 (m, 10H), 6.43–6.40 (m, 1H), 6.18 (s, 1H), 4.12 (q, *J* = 7.2 Hz, 2H), 4.00 (t, *J* = 5.5 Hz, 2H), 3.69 (s, 2H), 3.27 (s, 3H), 1.20 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 169.9, 169.7, 169.6, 138.1, 133.6, 130.6, 130.4, 129.0, 128.6, 128.6, 128.3, 70.8, 64.6, 61.3, 59.2, 41.6, 14.1.

(RS)-(E)-Ethyl 2-(2-phenyl-2-(*N*-phenylbut-2-enamido)acetamido)acetate (18c). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 g, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), crotonic acid (0.23 g, 1.00 mmol), and ethyl isocyanoacetate (0.11 mL, 1.00 mmol), to afford (18c) as a white solid (Method b 68% and c 51%); LRMS (ESI⁺) *m/z* 381 [M + H]⁺, mp 181.9–187.2; IR (cm⁻¹): 3331 (NH), 1662, 1625 (CO); ¹H NMR (400 MHz, CDCl₃) δ 7.15–7.25 (m, 9H), 6.91–7.02 (dd, *J* = 15.0, 6.9 Hz, 2H), 6.54–6.59 (s, 1H), 6.10–6.14 (s, 1H), 5.58–5.66 (dd, *J* = 15.1, 1.7 Hz, 1H), 4.14–4.21 (q, *J* = 7.2 Hz, 2H), 4.05–4.09 (dd, *J* = 5.3, 3.2 Hz, 2H), 1.67–1.72 (dd, *J* = 7.0, 1.7 Hz, 3H), 1.22–1.27 (t, *J* = 7.1 Hz, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 170.0, 169.8, 166.7, 142.8, 140.0, 134.3, 134.2, 130.5, 130.4, 129.0, 128.5, 128.4, 128.1, 122.8, 115.4, 65.7, 61.5, 41.7, 18.2, 14.2.

(RS)-Ethyl 2-(2-phenyl-2-(*N*-phenylbut-2-ynamido)acetyl)glycinate (18d). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), 2-butyric acid (0.23 g, 1.00 mmol), and ethyl isocyanoacetate (0.11 mL, 1.00 mmol), to afford (18d) as a white solid (Method b 66% and c 54%); LRMS (ESI⁺) *m/z* 379 [M + H]⁺, mp 175.6–178.4; IR (cm⁻¹): 3337 (NH), 1657, 1620 (CO), 1594, 1549; ¹H NMR (400 MHz, CDCl₃) δ 7.11–7.24 (m, 11H), 6.35 (s, 1H), 6.11 (s, 1H), 4.14–4.22 (q, *J* = 7.1 Hz, 2H), 4.03–4.10 (d, *J* = 5.3 Hz, 2H), 1.63–1.70 (s, 3H), 1.22–1.29 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 169.6, 169.3, 155.2, 139.7, 133.6, 130.6, 130.5, 128.8, 128.5, 128.5, 128.3, 91.7, 74.0, 64.9, 61.6, 60.5, 41.8, 21.1, 14.3, 14.2, 4.0.

(RS)-Ethyl 2-(2-(2-methoxy-*N*-phenylacetamido)-2-phenylacetyl)glycinate (18e). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 g, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), acetic acid (0.23 g, 2.70 mmol), and ethyl isocyanoacetate (0.11 mL, 1.00 mmol), to afford (18e) as a white solid (Method b 66% and c 69%); LRMS (ESI⁺) *m/z* 355 [M + H]⁺, mp 183.6–186.1. IR (cm⁻¹): 3244, 3085, 3061, 1655; ¹H NMR (400 MHz, CDCl₃) δ 6.97–7.18 (m, 10H), 6.59–6.65 (t, *J* = 5.4 Hz, 1H), 6.11–6.15 (s, 1H), 4.06–4.14 (q, *J* = 7.1 Hz, 2H), 3.95–4.01 (dd, *J* = 6.6, 5.4 Hz, 2H), 1.76–1.79 (s, 3H), 1.15–1.21 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 171.2, 170.0, 169.6, 140.5, 134.2, 130.5, 130.2, 128.8, 128.3, 128.2, 128.0, 64.6, 61.3, 41.5, 23.1, 14.1.

(RS)-Ethyl 2-(2-(2-cyano-*N*-phenylacetamido)-2-phenylacetamido)acetate (18f). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), cyanoacetic acid (0.23 g, 2.70 mmol), and ethyl isocyanoacetate (0.11 mL, 1.00 mmol), to afford (18f) as a white solid (Method b 58% and c 59%); LRMS (ESI⁺) *m/z* 380 [M + H]⁺, mp 168.2–172.2. IR (cm⁻¹): 3397 (NH), 2205 (CN) 1660, 1624 (CO); ¹H NMR (400 MHz, CDCl₃) δ 7.05–7.33 (m, 10H), 6.15–6.21 (s, 1H), 6.14 (s, 1H), 4.14–4.21 (q, *J* = 7.2 Hz, 2H), 4.01–4.10 (dd, *J* = 6.9, 5.3 Hz, 2H), 3.15–3.29 (m, 2H), 1.22–1.28 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 169.6, 169.1, 162.9, 138.5, 133.0, 130.8, 130.4, 129.4, 129.2, 128.8, 114.0, 65.6, 61.7, 41.8, 26.4, 14.2.

(RS)-Ethyl 2-(2-phenyl-2-(3,3,3-trifluoro-*N*-phenylpropanamido)acetyl)glycinate (18g). Synthesised using the general procedure (Method b and c) as described for (12) from aniline (0.09 mL, 1.00 mmol), benzaldehyde (0.10 g, 1.00 mmol), 3,3,3-trifluoropropionic acid (0.23 g, 1.00 mmol), and ethyl isocyanoacetate (0.11 mL, 1.00 mmol), to afford (18g) as a white solid (Method b 50% and c 45%); LRMS (ESI⁺) *m/z* 423 [M + H]⁺, mp 164.2–166.5. IR (cm⁻¹): 3393 (NH), 2206 (CN) 1660, 1624 (CO); ¹H NMR (400 MHz, CDCl₃): δ 7.56–7.74 (s, 1H), 7.02–7.42 (m, 9H), 6.46–6.64 (d, *J* = 7.1 Hz, 1H), 6.19–6.25 (s, 1H), 6.12–6.16 (s, 1H), 4.13–4.21 (q, *J* = 7.1 Hz, 2H), 4.03–4.08 (dd, *J* = 5.3, 2.7 Hz, 2H), 2.83–3.03 (qd, *J* = 10.0, 7.9 Hz, 2H), 1.21–1.27 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃): δ 169.6, 169.3, 164.2, 164.2, 138.9, 133.4, 130.7, 130.5, 129.0, 128.7, 65.2, 61.7, 41.8, 39.5, 39.2, 38.9, 38.6, 14.2.

(RS)-*N*-(2-(Cyclohexylamino)-2-oxo-1-phenylethyl)-*N*-(4-methoxyphenyl)propiolamide (19). Synthesised using the general procedure (method a) from 4-methoxybenzaldehyde (0.136 g, 1.00 mmol), 2-butyric acid (0.23 g, 1.00 mmol), aniline (0.9 mL, 1.00 mmol) and cyclohexyl isocyanide (0.11 mL, 1.00 mmol) to afford (19) as a white solid (0.259 g, 66%) LRMS *m/z* 391 [M + H]⁺ mp: 169–172; ¹H NMR (400 MHz, CDCl₃) δ 7.24–7.20 (m, 3H, signal overlaps residual CHCl₃), 7.17–7.16 (m, 2H), 7.09 (t, *J* = 8.1 Hz, 1H), 6.79 (dd, *J* = 8.2, 2.2 Hz, 2H), 6.74 (bs s, 1H), 5.96 (s, 1H), 5.53 (d, *J* = 7.7 Hz, 1H), 3.87–3.78 (m, 1H), 3.66, s, 3H), 2.82 (s, 1H) 1.89 (dd, *J* = 34.5, 10.4 Hz, 2H), 1.68–1.56 (m, 3H, signal overlaps residual water), 1.40–1.28 (m, 2H), 1.17–0.98 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 167.7, 159.7, 153.8, 140.3, 134.0, 130.4 (2C), 129.2, 128.9, 128.7 (2C), 123.2, 115.8, 115.1, 80.5, 76.2, 65.4, 55.5, 49.0, 32.95, 32.90, 25.6, 24.9, 24.8.

(RS)-*N*-(2-(Cyclohexylamino)-1-(2,3-dimethoxyphenyl)-2-oxoethyl)-*N*-phenylpropiolamide (20). Synthesised by method A from 2,3-dimethoxybenzaldehyde (0.166 g, 1.00 mmol), 2-butyric acid (0.23 g, 1.00 mmol), aniline (0.9 mL, 1.00 mmol) and cyclohexyl isocyanide (0.11 mL, 1.00 mmol) to afford (20) as a white solid (0.2682 g, 64%). LRMS *m/z* 421.2 [M + H]⁺ mp 224 (dec); ¹H NMR (400 MHz, CDCl₃) δ 7.24–7.17 (m, 5H), 6.82–6.74 (m, 2H), 6.59 (dd, *J* = 7.4, 1.4 Hz, 1H), 6.40 (s, 1H), 5.49 (d, *J* = 7.5 Hz, 1H), 3.84–3.77 (m, 7H), 2.78 (s, 1H), 1.89 (dd, *J* = 38.5, 10.8 Hz, 2H), 1.70–1.55 (m, 3H, signal overlaps residual water), 1.40–1.27 (m, 2H), 1.16–0.97 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.1, 153.8, 152.5, 147.9, 139.5, 130.7 (2C), 128.59, 128.56 (2C), 127.8, 123.7, 122.8, 113.2, 80.4, 76.4, 61.1, 58.8, 55.9, 49.0, 33.0, 32.90, 25.6, 25.0, 24.9.

(RS)-*N*-(2-(Cyclohexylamino)-1-(3,4-dimethoxyphenyl)-2-oxoethyl)-*N*-phenylpropiolamide (21). Synthesised by method A from 2,3-dimethoxybenzaldehyde (0.166 g, 1.00 mmol), 2-butyric acid (0.23 g, 1.00 mmol), aniline (0.9 mL, 1.00 mmol) and cyclohexyl isocyanide (0.11 mL, 1.00 mmol) to afford (21) as a white solid (0.231 g, 55%). LRMS *m/z* 421.2 [M + H]⁺ ¹H NMR (400 MHz, CDCl₃) δ 7.24–7.17 (m, 5H, overlapping signal with CDCl₃), 6.77–6.69 (m, 2H), 6.51 (s, 1H), 6.00 (s, 1H), 5.57 (d, *J* = 7.6 Hz, 1H), 3.83 (s, 3H) 3.81–3.75 (m, 1H), 3.60 (s, 3H), 2.80 (s, 1H), 1.88 (dd, *J* = 48.3, 11.2 Hz, 2H), 1.69–1.56 (m, 3H, overlapping signal with residual water), 1.39–1.28 (m, 2H), 1.16–0.98



(m, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 168.0, 153.9, 149.44, 148.7, 139.0, 131.10 (2C), 128.69, 128.66 (2C), 126.1, 123.4 (2C), 113.4, 110.7, 80.7, 64.3, 55.9, 55.8, 49.0, 32.9 (2C), 25.6, 24.9, 24.8.

Conflicts of interest

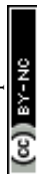
The authors declare no conflict of interest.

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